# Package 'nandb'

May 17, 2021

```
Type Package
Title Number and Brightness Image Analysis
Version 2.1.0
Description Calculation of molecular number and brightness from
     fluorescence microscopy image series. The software was published in a
     2016 paper <doi:10.1093/bioinformatics/btx434>. The seminal paper for
     the technique is Digman et al. 2008 <doi:10.1529/biophysj.107.114645>.
     A review of the technique was published in 2017
     <doi:10.1016/j.ymeth.2017.12.001>.
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URL https://rorynolan.github.io/nandb/,
     https://github.com/rorynolan/nandb
BugReports https://github.com/rorynolan/nandb/issues
Depends R (>= 3.1)
Imports assertthat, autothresholdr (>= 1.3.11), BBmisc, checkmate (>=
     1.9.3), detrendr (>= 0.6.12), dplyr, filesstrings (>= 3.2),
     ggplot2, glue (>= 1.3), ijtiff (>= 2.2), magrittr (>= 1.5),
     purrr, Rcpp (>= 1.0.1), reshape2, rlang (>= 0.3.3), stringr (>=
     1.4), utils, viridis, with (>= 2.1.0)
Suggests abind, covr, gridExtra, knitr, matrixStats (>= 0.50), pacman,
     rmarkdown, spelling, testthat (>= 2.1), tidyr
LinkingTo Rcpp (>= 1.0.1)
VignetteBuilder knitr
Config/testthat/edition 3
Config/testthat/parallel true
Encoding UTF-8
Language en-US
RoxygenNote 7.1.1
NeedsCompilation yes
```

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## Repository CRAN

**Date/Publication** 2021-05-16 22:40:06 UTC

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		_
brig	htness Calculate brightness from image series.	

## Description

Given a time stack of images, brightness() performs a calculation of the brightness for each pixel.

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### Usage

```
brightness(
  img,
  def,
  thresh = NULL,
  detrend = FALSE,
  quick = FALSE,
  filt = NULL,
  s = 1,
  offset = 0,
  readout_noise = 0,
  parallel = FALSE
)
```

#### **Arguments**

detrend

filt

img	A 4-dimensional array in the style of an ijtiff_img (indexed by img[y,x,channel,frame]) or a 3-dimensional array which is a single channel of an ijtiff_img (indexed by img[y,x,frame]).
def	A character. Which definition of brightness do you want to use, "B" or "epsilon"?
thresh	The threshold or thresholding method (see autothresholdr::mean_stack_thresh())

to use on the image prior to detrending and brightness calculations.

Detrend your data with detrendr::img\_detrend\_rh(). This is the best known detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. If there are many channels, this

may be specified as a vector, one element for each channel.

quick If FALSE (the default), the swap finding routine is run several times to get a consensus for the best parameter. If TRUE, the swap finding routine is run only

once.

Do you want to smooth (filt = 'mean') or median (filt = 'median') filter the number image using smooth\_filter() or median\_filter() respectively? If selected, these are invoked here with a filter radius of 1 (with corners included, so each median is the median of 9 elements) and with the option na\_count =

TRUE. If you want to smooth/median filter the number image in a different way, first calculate the numbers without filtering (filt = NULL) using this function and then perform your desired filtering routine on the result. If there are many

channels, this may be specified as a vector, one element for each channel.

s A positive number. The S-factor of microscope acquisition. offset Microscope acquisition parameters. See reference Dalal et al. Microscope acquisition parameters. See reference Dalal et al. readout\_noise

parallel Would you like to use multiple cores to speed up this function? If so, set the

number of cores here, or to use all available cores, use parallel = TRUE.

### Value

A matrix, the brightness image.

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#### References

Digman MA, Dalal R, Horwitz AF, Gratton E. Mapping the Number of Molecules and Brightness in the Laser Scanning Microscope. Biophysical Journal. 2008;94(6):2320-2332. doi: 10.1529/biophysj.107.114645.

Dalal, RB, Digman, MA, Horwitz, AF, Vetri, V, Gratton, E (2008). Determination of particle number and brightness using a laser scanning confocal microscope operating in the analog mode. Microsc. Res. Tech., 71, 1:69-81. doi: 10.1002/jemt.20526.

## **Examples**

```
img <- ijtiff::read_tif(system.file("extdata", "50.tif", package = "nandb"))
ijtiff::display(img[, , 1, 1])
b <- brightness(img, "e", thresh = "Huang")
b <- brightness(img, "B", thresh = "tri")</pre>
```

brightness\_folder

Brightness calculations for every image in a folder.

## **Description**

Perform brightness() calculations on all tif images in a folder and save the resulting brightness images to disk.

#### Usage

```
brightness_folder(
  folder_path = ".",
  def,
  thresh = NULL,
  detrend = FALSE,
  quick = FALSE,
  filt = NULL,
  s = 1,
  offset = 0,
  readout_noise = 0,
  parallel = FALSE
)
```

## **Arguments**

folder\_path The path (relative or absolute) to the folder you wish to process.

def A character. Which definition of brightness do you want to use, "B" or "epsilon"?

thresh The threshold or thresholding method (see autothresholdr::mean\_stack\_thresh()) to use on the image prior to detrending and brightness calculations.

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detrend Detrend your data with detrendr::img\_detrend\_rh(). This is the best known detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. If there are many channels, this may be specified as a vector, one element for each channel. quick If FALSE (the default), the swap finding routine is run several times to get a consensus for the best parameter. If TRUE, the swap finding routine is run only filt Do you want to smooth (filt = 'mean') or median (filt = 'median') filter the number image using smooth\_filter() or median\_filter() respectively? If selected, these are invoked here with a filter radius of 1 (with corners included, so each median is the median of 9 elements) and with the option na\_count = TRUE. If you want to smooth/median filter the number image in a different way, first calculate the numbers without filtering (filt = NULL) using this function and then perform your desired filtering routine on the result. If there are many channels, this may be specified as a vector, one element for each channel. s A positive number. The S-factor of microscope acquisition. offset Microscope acquisition parameters. See reference Dalal et al. readout\_noise Microscope acquisition parameters. See reference Dalal et al. parallel Would you like to use multiple cores to speed up this function? If so, set the number of cores here, or to use all available cores, use parallel = TRUE.

#### See Also

number()

### **Examples**

```
## Not run:
setwd(tempdir())
img <- ijtiff::read_tif(system.file("extdata", "50.tif", package = "nandb"))
ijtiff::write_tif(img, "img1.tif")
ijtiff::write_tif(img, "img2.tif")
brightness_folder(def = "B", thresh = "Huang")
## End(Not run)</pre>
```

### **Description**

Given a stack of images img, use the first frames\_per\_set of them to create one brightness image, the next frames\_per\_set of them to create the next brightness image and so on to get a time-series of brightness images.

#### Usage

```
brightness_timeseries(
  img,
  def,
  frames_per_set,
  overlap = FALSE,
  thresh = NULL,
  detrend = FALSE,
  quick = FALSE,
  filt = NULL,
  s = 1,
  offset = 0,
  readout_noise = 0,
  parallel = FALSE
)
```

#### **Arguments**

s

img A 4-dimensional array in the style of an ijtiff\_img (indexed by img[y,x,channel,frame])

or a 3-dimensional array which is a single channel of an ijtiff\_img (indexed by

img[y,x,frame]).

def A character. Which definition of brightness do you want to use, "B" or "epsilon"?

frames\_per\_set The number of frames with which to calculate the successive brightnesses.

overlap A boolean. If TRUE, the windows used to calculate number are overlapped, if

FALSE, they are not. For example, for a 20-frame image series with 5 frames per set, if the windows are not overlapped, then the frame sets used are 1-5, 6-10, 11-15 and 16-20; whereas if they are overlapped, the frame sets are 1-5, 2-6,

3-7, 4-8 and so on up to 16-20.

thresh The threshold or thresholding method (see autothresholdr::mean\_stack\_thresh())

to use on the image prior to detrending and brightness calculations.

detrend Detrend your data with detrendr::img\_detrend\_rh(). This is the best known

detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. If there are many channels, this

may be specified as a vector, one element for each channel.

quick If FALSE (the default), the swap finding routine is run several times to get a

consensus for the best parameter. If TRUE, the swap finding routine is run only

once.

filt Do you want to smooth (filt = 'mean') or median (filt = 'median') filter the

number image using smooth\_filter() or median\_filter() respectively? If selected, these are invoked here with a filter radius of 1 (with corners included, so each median is the median of 9 elements) and with the option na\_count = TRUE. If you want to smooth/median filter the number image in a different way, first calculate the numbers without filtering (filt = NULL) using this function and then perform your desired filtering routine on the result. If there are many

channels, this may be specified as a vector, one element for each channel.

A positive number. The S-factor of microscope acquisition.

offset Microscope acquisition parameters. See reference Dalal et al. readout\_noise Microscope acquisition parameters. See reference Dalal et al.

parallel Would you like to use multiple cores to speed up this function? If so, set the

number of cores here, or to use all available cores, use parallel = TRUE.

#### **Details**

This may discard some images, for example if 175 frames are in the input and frames\_per\_set = 50, then the last 25 are discarded. If detrending is selected, it is performed on the whole image stack before the sectioning is done for calculation of numbers.

#### Value

An object of class brightness\_ts\_img.

- If img is 3-dimensional (i.e. 1-channel), a 3-dimensional array arr is returned with arr[y, x, t] being pixel (x, y) of the tth brightness image in the brightness time series.
- If img is 4-dimensional (i.e. 2-channel), a 4-dimensional array arr is returned with arr[y,x,c,t] being pixel (x,y) of the cth channel of the tth brightness image in the brightness time series.

#### See Also

```
brightness().
```

#### **Examples**

```
img <- ijtiff::read_tif(system.file("extdata", "50.tif", package = "nandb"))
bts <- brightness_timeseries(img, "e", frames_per_set = 20, thresh = "Huang")</pre>
```

```
brightness_timeseries_folder
```

Brightness time-series calculations for every image in a folder.

#### **Description**

Perform brightness\_timeseries() calculations on all tif images in a folder and save the resulting number images to disk.

#### Usage

```
brightness_timeseries_folder(
  folder_path = ".",
  def,
  frames_per_set,
  overlap = FALSE,
  thresh = NULL,
  detrend = FALSE,
  quick = FALSE,
  filt = NULL,
  s = 1,
  offset = 0,
  readout_noise = 0,
  parallel = FALSE
)
```

## **Arguments**

folder\_path The path (relative or absolute) to the folder you wish to process.

def A character. Which definition of brightness do you want to use, "B" or "epsilon"?

frames\_per\_set The number of frames with which to calculate the successive brightnesses.

overlap A boolean. If TRUE, the windows used to calculate number are overlapped, if

FALSE, they are not. For example, for a 20-frame image series with 5 frames per set, if the windows are not overlapped, then the frame sets used are 1-5, 6-10, 11-15 and 16-20; whereas if they are overlapped, the frame sets are 1-5, 2-6,

3-7, 4-8 and so on up to 16-20.

thresh The threshold or thresholding method (see autothresholdr::mean\_stack\_thresh())

to use on the image prior to detrending and brightness calculations.

detrend Detrend your data with detrendr::img\_detrend\_rh(). This is the best known

detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. If there are many channels, this

may be specified as a vector, one element for each channel.

quick If FALSE (the default), the swap finding routine is run several times to get a

consensus for the best parameter. If TRUE, the swap finding routine is run only

once.

filt Do you want to smooth (filt = 'mean') or median (filt = 'median') filter the

number image using smooth\_filter() or median\_filter() respectively? If selected, these are invoked here with a filter radius of 1 (with corners included, so each median is the median of 9 elements) and with the option na\_count = TRUE. If you want to smooth/median filter the number image in a different way, first calculate the numbers without filtering (filt = NULL) using this function and then perform your desired filtering routine on the result. If there are many

channels, this may be specified as a vector, one element for each channel.

s A positive number. The S-factor of microscope acquisition.

offset Microscope acquisition parameters. See reference Dalal et al.

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readout\_noise Microscope acquisition parameters. See reference Dalal et al.

parallel Would you like to use multiple cores to speed up this function? If so, set the

number of cores here, or to use all available cores, use parallel = TRUE.

#### See Also

```
brightness_timeseries()
```

### **Examples**

```
## Not run:
setwd(tempdir())
img <- ijtiff::read_tif(system.file("extdata", "50.tif", package = "nandb"))
ijtiff::write_tif(img, "img1.tif")
ijtiff::write_tif(img, "img2.tif")
brightness_timeseries_folder(def = "e", thresh = "tri", frames_per_set = 20)
## End(Not run)</pre>
```

cc-nb-img-classes

Cross-correlated number and brightness image classes.

#### **Description**

The cc\_number\_img and cc\_brightness\_img classes are designed to hold objects which are images calculated from the *cross-correlated number and brightness* technique.

#### Usage

```
cc_number_img(img, thresh, swaps, filt)
cc_brightness_img(img, thresh, swaps, filt)
```

## **Arguments**

img The calculated cross-correlated number or brightness image.

thresh A positive integer, possibly an object of class autothresholdr::th. If the different

channels of the image had different thresholds, this argument may be specified as a vector or list (of positive integers, possibly objects of class autothresholdr::th),

one element for each channel.

swaps A non-negative integer with an attribute auto. If the different channels of the im-

age had different swaps, this argument may be specified as a list (of non-negative integers with attributes auto), one element for each channel. For undetrended

images, set swaps = NA.

filt A string, the filtering method used. Must be either "mean" or "median", or NA

for no filtering. If the different channels of the image had different filters, this

may be specified as a character vector, one element for each channel.

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#### **Details**

An object of class cc\_number\_img or cc\_brightness\_img is a 4-dimensional array of real numbers in the mould of an ijtiff\_img (indexed as img[y,x,channel,frame]) with 3 attributes:

thresh A positive integer, possibly an object of class autothresholdr::th detailing which threshold and thresholding method was used in preprocessing (in the multi-channel case, one threshold per channel is given).

swaps A non-negative integer indicating the number of swaps used for Robin Hood detrending, with an attribute auto which is a logical indicating whether or not the parameter was chosen automatically (in the multi-channel case, one swaps per channel is given).

filt Was mean or median filtering used in postprocessing?

#### Value

An object of class cc\_number\_img or cc\_brightness\_img.

cc-nb-ts-img-classes Cross-correlated number and brightness time series image classes.

## **Description**

The cc\_number\_ts\_img and cc\_brightness\_ts\_img classes are designed to hold objects which are images calculated from the *cross-correlated number and brightness* technique.

## Usage

```
cc_number_ts_img(img, frames_per_set, overlapped, thresh, swaps, filt)
cc_brightness_ts_img(img, frames_per_set, overlapped, thresh, swaps, filt)
```

## **Arguments**

img	The calculated cross-correlated number or brightness time series image series.
frames_per_set	The number of frames used in the calculation of each point in the cross-correlated number or brightness time series.
overlapped	A boolean. TRUE indicates that the windows used to calculate consecutive brightnesses over time were overlapped, FALSE indicates that they were not.
thresh	A positive integer, possibly an object of class autothresholdr::th. If the different channels of the image had different thresholds, this argument may be specified as a vector or list (of positive integers, possibly objects of class autothresholdr::th), one element for each channel.
swaps	A non-negative integer with an attribute auto. If the different channels of the image had different swaps, this argument may be specified as a list (of non-negative integers with attributes auto), one element for each channel. For undetrended images, set swaps = NA.
filt	A string, the filtering method used. Must be either "mean" or "median", or NA for no filtering. If the different channels of the image had different filters, this may be specified as a character vector, one element for each channel.

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#### **Details**

An object of class cc\_number\_ts\_img or cc\_brightness\_ts\_img is a 4-dimensional array of real numbers in the mould of an ijtiff\_img with 3 attributes:

thresh A positive integer, possibly an object of class autothresholdr::th detailing which threshold and thresholding method was used in preprocessing (in the multi-channel case, one threshold per channel is given).

swaps A non-negative integer indicating the parameter used for Robin Hood detrending with an attribute auto which is a logical indicating whether or not the parameter was chosen automatically (in the multi-channel case, one swaps per channel is given).

frames\_per\_set A positive integer detailing how many frames were used in the calculation of each point in the number or brightness time series.

overlapped A boolean. TRUE indicates that the windows used to calculate consecutive brightnesses over time were overlapped, FALSE indicates that they were not.

#### Value

An object of class cc\_number\_ts\_img or cc\_brightness\_ts\_img.

#### See Also

```
cc_number_timeseries(), cc_brightness_timeseries().
```

cc\_brightness

Cross-correlated brightness.

### Description

Given a time stack of images and two channels, calculate the cross-correlated brightness of those two channels for each pixel.

```
cc_brightness(
  img,
  ch1 = 1,
  ch2 = 2,
  thresh = NULL,
  detrend = FALSE,
  quick = FALSE,
  filt = NULL,
  parallel = FALSE
)
```

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#### **Arguments**

img A 4-dimensional array of images indexed by img[y,x,channel,frame] (an object of class ijtiff::ijtiff\_img). The image to perform the calculation on. To perform this on a file that has not yet been read in, set this argument to the path to that file (a string). ch1 A natural number. The index of the first channel to use. ch2 A natural number. The index of the second channel to use. thresh Do you want to apply an intensity threshold prior to calculating cross-correlated brightness (via autothresholdr::mean\_stack\_thresh())? If so, set your thresholding method here. If this is a single value, that same threshold will be applied to both channels. If this is a length-2 vector or list, then these two thresholds will be applied to channels 1 and 2 respectively. A value of NA for either channel gives no thresholding for that channel. detrend Detrend your data with detrendr::img\_detrend\_rh(). This is the best known detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. To detrend one channel and not the other, specify this as a length 2 vector. quick FALSE repeats the detrending procedure (which has some inherent randomness) a few times to hone in on the best detrend. TRUE is quicker, performing the routine only once. FALSE is better. filt Do you want to smooth (filt = 'smooth') or median (filt = 'median') filter the cross-correlated brightness image using smooth\_filter() or median\_filter() respectively? If selected, these are invoked here with a filter radius of 1 and with the option na\_count = TRUE. A value of NA for either channel gives no thresholding for that channel. If you want to smooth/median filter the cross-correlated brightness image in a different way, first calculate the cross-correlated brightnesses without filtering (filt = NULL) using this function and then perform your desired filtering routine on the result. parallel Would you like to use multiple cores to speed up this function? If so, set the number of cores here, or to use all available cores, use parallel = TRUE.

#### Value

A numeric matrix, the cross-correlated brightness image.

#### **Examples**

```
img <- ijtiff::read_tif(system.file("extdata", "two_ch.tif",
    package = "nandb"
))
ijtiff::display(detrendr::mean_pillars(img[, , 1, ]))
ijtiff::display(detrendr::mean_pillars(img[, , 2, ]))
b <- brightness(img, def = "e", thresh = "Huang", filt = "median")
ijtiff::display(b[, , 1, 1])
ijtiff::display(b[, , 2, 1])
cc_b <- cc_brightness(img, thresh = "Huang")</pre>
```

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```
ijtiff::display(cc_b[, , 1, 1])
```

 $\verb|cc_brightness_folder| Cross-correlated brightness calculations for every image in a folder.$ 

## Description

Perform cc\_brightness() calculations on all TIFF images in a folder and save the resulting images to disk.

## Usage

```
cc_brightness_folder(
  folder_path = ".",
  ch1 = 1,
  ch2 = 2,
  thresh = NULL,
  detrend = detrend,
  quick = quick,
  filt = NULL,
  parallel = FALSE
)
```

## **Arguments**

folder_path	The path (relative or absolute) to the folder you wish to process.
ch1	A natural number. The index of the first channel to use.
ch2	A natural number. The index of the second channel to use.
thresh	Do you want to apply an intensity threshold prior to calculating cross-correlated brightness (via autothresholdr::mean_stack_thresh())? If so, set your thresholding method here. If this is a single value, that same threshold will be applied to both channels. If this is a length-2 vector or list, then these two thresholds will be applied to channels 1 and 2 respectively. A value of NA for either channel gives no thresholding for that channel.
detrend	Detrend your data with detrendr::img_detrend_rh(). This is the best known detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. To detrend one channel and not the other, specify this as a length 2 vector.
quick	FALSE repeats the detrending procedure (which has some inherent randomness) a few times to hone in on the best detrend. TRUE is quicker, performing the routine only once. FALSE is better.
filt	Do you want to smooth (filt = 'smooth') or median (filt = 'median') filter the cross-correlated brightness image using smooth_filter() or median_filter() respectively? If selected, these are invoked here with a filter radius of 1 and with

the option na\_count = TRUE. A value of NA for either channel gives no thresholding for that channel. If you want to smooth/median filter the cross-correlated brightness image in a different way, first calculate the cross-correlated brightnesses without filtering (filt = NULL) using this function and then perform your desired filtering routine on the result.

parallel

Would you like to use multiple cores to speed up this function? If so, set the number of cores here, or to use all available cores, use parallel = TRUE.

#### **Examples**

```
## Not run:
setwd(tempdir())
ijtiff::write_tif(img, "a.tif")
ijtiff::write_tif(img, "ab.tif")
cc_brightness_folder()
list.files()
## End(Not run)
```

cc\_brightness\_timeseries

*Create a cross-correlated brightness time-series.* 

### Description

Given a stack of images img, use the first frames\_per\_set of them to create one cross-correlated brightness image, the next frames\_per\_set of them to create the next and so on to get a time-series of cross-correlated brightness images.

```
cc_brightness_timeseries(
  img,
  frames_per_set,
  overlap = FALSE,
  ch1 = 1,
  ch2 = 2,
  thresh = NULL,
  detrend = FALSE,
  quick = FALSE,
  filt = NULL,
  parallel = FALSE
)
```

#### **Arguments**

img

A 4-dimensional array of images indexed by img[y,x,channel,frame] (an object of class ijtiff::ijtiff\_img). The image to perform the calculation on. To perform this on a file that has not yet been read in, set this argument to the path to that file (a string).

frames\_per\_set The number of frames with which to calculate the successive cross-correlated brightnesses.

> This may discard some images, for example if 175 frames are in the input and frames\_per\_set = 50, then the last 25 are discarded. If bleaching or/and thresholding are selected, they are performed on the whole image stack before the sectioning is done for calculation of cross-correlated brightnesses.

overlap

A boolean. If TRUE, the windows used to calculate brightness are overlapped, if FALSE, they are not. For example, for a 20-frame image series with 5 frames per set, if the windows are not overlapped, then the frame sets used are 1-5, 6-10, 11-15 and 16-20; whereas if they are overlapped, the frame sets are 1-5, 2-6, 3-7, 4-8 and so on up to 16-20.

ch1 A natural number. The index of the first channel to use.

ch2 A natural number. The index of the second channel to use.

thresh Do you want to apply an intensity threshold prior to calculating cross-correlated

> brightness (via autothresholdr::mean\_stack\_thresh())? If so, set your thresholding method here. If this is a single value, that same threshold will be applied to both channels. If this is a length-2 vector or list, then these two thresholds will be applied to channels 1 and 2 respectively. A value of NA for

either channel gives no thresholding for that channel.

detrend Detrend your data with detrendr::img\_detrend\_rh(). This is the best known

> detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. To detrend one channel and not the

other, specify this as a length 2 vector.

FALSE repeats the detrending procedure (which has some inherent randomness)

a few times to hone in on the best detrend. TRUE is quicker, performing the

routine only once. FALSE is better.

filt Do you want to smooth (filt = 'smooth') or median (filt = 'median') filter

> the cross-correlated brightness image using smooth\_filter() or median\_filter() respectively? If selected, these are invoked here with a filter radius of 1 and with the option na\_count = TRUE. A value of NA for either channel gives no thresholding for that channel. If you want to smooth/median filter the cross-correlated brightness image in a different way, first calculate the cross-correlated brightnesses without filtering (filt = NULL) using this function and then perform your

desired filtering routine on the result.

Would you like to use multiple cores to speed up this function? If so, set the parallel

number of cores here, or to use all available cores, use parallel = TRUE.

### Value

An array where the *i*th slice is the *i*th cross-correlated brightness image.

quick

#### See Also

```
brightness().
```

### **Examples**

```
img <- ijtiff::read_tif(system.file("extdata", "two_ch.tif",
   package = "nandb"
))
cc_bts <- cc_brightness_timeseries(img, 10,
   thresh = "Huang",
   filt = "median", parallel = 2
)
ijtiff::display(cc_bts[, , 1, 1])</pre>
```

cc\_brightness\_timeseries\_folder

Cross-correlated brightness time-series calculations for every image in a folder.

## Description

Perform cc\_brightness\_timeseries() calculations on all tif images in a folder and save the resulting images to disk.

## Usage

```
cc_brightness_timeseries_folder(
  folder_path = ".",
  frames_per_set,
  overlap = FALSE,
  ch1 = 1,
  ch2 = 2,
  thresh = NULL,
  detrend = detrend,
  quick = quick,
  filt = NULL,
  parallel = FALSE
)
```

### **Arguments**

folder\_path The path (relative or absolute) to the folder you wish to process.

frames\_per\_set The number of frames with which to calculate the successive cross-correlated brightnesses.

This may discard some images, for example if 175 frames are in the input and frames\_per\_set = 50, then the last 25 are discarded. If bleaching or/and thresholding are selected, they are performed on the whole image stack before the sectioning is done for calculation of cross-correlated brightnesses.

overlap

A boolean. If TRUE, the windows used to calculate brightness are overlapped, if FALSE, they are not. For example, for a 20-frame image series with 5 frames per set, if the windows are not overlapped, then the frame sets used are 1-5, 6-10, 11-15 and 16-20; whereas if they are overlapped, the frame sets are 1-5, 2-6, 3-7, 4-8 and so on up to 16-20.

ch1 A natural number. The index of the first channel to use.

ch2 A natural number. The index of the second channel to use.

thresh Do you want to apply an intensity threshold prior to calculating cross-correlated

brightness (via autothresholdr::mean\_stack\_thresh())? If so, set your thresholding method here. If this is a single value, that same threshold will be applied to both channels. If this is a length-2 vector or list, then these two thresholds will be applied to channels 1 and 2 respectively. A value of NA for

either channel gives no thresholding for that channel.

detrend Detrend your data with detrendr::img\_detrend\_rh(). This is the best known

detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. To detrend one channel and not the

other, specify this as a length 2 vector.

quick FALSE repeats the detrending procedure (which has some inherent randomness)

a few times to hone in on the best detrend. TRUE is quicker, performing the

routine only once. FALSE is better.

filt Do you want to smooth (filt = 'smooth') or median (filt = 'median') filter

the cross-correlated brightness image using smooth\_filter() or median\_filter() respectively? If selected, these are invoked here with a filter radius of 1 and with the option na\_count = TRUE. A value of NA for either channel gives no thresholding for that channel. If you want to smooth/median filter the cross-correlated brightness image in a different way, first calculate the cross-correlated brightnesses without filtering (filt = NULL) using this function and then perform your

desired filtering routine on the result.

parallel Would you like to use multiple cores to speed up this function? If so, set the

number of cores here, or to use all available cores, use parallel = TRUE.

### See Also

```
cc_brightness_timeseries()
```

### **Examples**

```
## Not run:
setwd(tempdir())
ijtiff::write_tif(img, "a.tif")
ijtiff::write_tif(img, "ab.tif")
cc_brightness_timeseries_folder(frames_per_set = 25)
list.files()
```

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```
## End(Not run)
```

cc\_number

Cross-correlated number.

#### **Description**

Given a time stack of images and two channels, calculate the cross-correlated number of those two channels for each pixel.

## Usage

```
cc_number(
  img,
  ch1 = 1,
  ch2 = 2,
  thresh = NULL,
  detrend = FALSE,
  quick = FALSE,
  filt = NULL,
  parallel = FALSE
)
```

#### **Arguments**

img	A 4-dimensional array of images indexed by img[y,x,channel,frame] (an ob-
-----	---

ject of class ijtiff::ijtiff\_img). The image to perform the calculation on. To perform this on a file that has not yet been read in, set this argument to the path to

that file (a string).

ch1 A natural number. The index of the first channel to use.

ch2 A natural number. The index of the second channel to use.

thresh Do you want to apply an intensity threshold prior to calculating cross-correlated

number (via autothresholdr::mean\_stack\_thresh())? If so, set your thresholding method here. If this is a single value, that same threshold will be applied to both channels. If this is a length-2 vector or list, then these two thresholds will be applied to channels 1 and 2 respectively. A value of NA for either channel

gives no thresholding for that channel.

detrend Detrend your data with detrendr::img\_detrend\_rh(). This is the best known

detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. To detrend one channel and not the

other, specify this as a length 2 vector.

quick FALSE repeats the detrending procedure (which has some inherent randomness)

a few times to hone in on the best detrend. TRUE is quicker, performing the

routine only once. FALSE is better.

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filt

Do you want to smooth (filt = 'smooth') or median (filt = 'median') filter the cross-correlated number image using smooth\_filter() or median\_filter() respectively? If selected, these are invoked here with a filter radius of 1 and with the option na\_count = TRUE. A value of NA for either channel gives no thresholding for that channel. If you want to smooth/median filter the cross-correlated number image in a different way, first calculate the cross-correlated numbers without filtering (filt = NULL) using this function and then perform your desired filtering routine on the result.

parallel

Would you like to use multiple cores to speed up this function? If so, set the number of cores here, or to use all available cores, use parallel = TRUE.

#### Value

A numeric matrix, the cross-correlated number image.

#### **Examples**

```
img <- ijtiff::read_tif(system.file("extdata", "two_ch.tif",
    package = "nandb"
))
ijtiff::display(detrendr::mean_pillars(img[, , 1, ]))
ijtiff::display(detrendr::mean_pillars(img[, , 2, ]))
n <- number(img, def = "n", thresh = "Huang", filt = "median")
ijtiff::display(n[, , 1, 1])
ijtiff::display(n[, , 2, 1])
cc_n <- cc_number(img, thresh = "Huang")
ijtiff::display(cc_n[, , 1, 1])</pre>
```

cc\_number\_folder

*Cross-correlated number calculations for every image in a folder.* 

## **Description**

Perform cc\_number() calculations on all TIFF images in a folder and save the resulting images to disk.

```
cc_number_folder(
  folder_path = ".",
  ch1 = 1,
  ch2 = 2,
  thresh = NULL,
  detrend = FALSE,
  quick = FALSE,
  filt = NULL,
  parallel = FALSE
)
```

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#### **Arguments**

folder\_path The path (relative or absolute) to the folder you wish to process.

ch1 A natural number. The index of the first channel to use.ch2 A natural number. The index of the second channel to use.

thresh Do you want to apply an intensity threshold prior to calculating cross-correlated

number (via autothresholdr::mean\_stack\_thresh())? If so, set your thresholding method here. If this is a single value, that same threshold will be applied to both channels. If this is a length-2 vector or list, then these two thresholds will be applied to channels 1 and 2 respectively. A value of NA for either channel

gives no thresholding for that channel.

detrend Detrend your data with detrendr::img\_detrend\_rh(). This is the best known

detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. To detrend one channel and not the

other, specify this as a length 2 vector.

quick FALSE repeats the detrending procedure (which has some inherent randomness)

a few times to hone in on the best detrend. TRUE is quicker, performing the

routine only once. FALSE is better.

filt Do you want to smooth (filt = 'smooth') or median (filt = 'median') filter

the cross-correlated number image using smooth\_filter() or median\_filter() respectively? If selected, these are invoked here with a filter radius of 1 and with the option na\_count = TRUE. A value of NA for either channel gives no thresholding for that channel. If you want to smooth/median filter the cross-correlated number image in a different way, first calculate the cross-correlated numbers without filtering (filt = NULL) using this function and then perform your de-

sired filtering routine on the result.

parallel Would you like to use multiple cores to speed up this function? If so, set the

number of cores here, or to use all available cores, use parallel = TRUE.

## **Examples**

```
## Not run:
setwd(tempdir())
ijtiff::write_tif(img, "a.tif")
ijtiff::write_tif(img, "ab.tif")
cc_number_folder()
list.files()
## End(Not run)
```

#### **Description**

Given a stack of images img, use the first frames\_per\_set of them to create one cross-correlated number image, the next frames\_per\_set of them to create the next and so on to get a time-series of cross-correlated number images.

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#### Usage

```
cc_number_timeseries(
  img,
  frames_per_set,
  overlap = FALSE,
  ch1 = 1,
  ch2 = 2,
  thresh = NULL,
  detrend = FALSE,
  quick = FALSE,
  filt = NULL,
  parallel = FALSE
)
```

## **Arguments**

img

A 4-dimensional array of images indexed by img[y,x,channel,frame] (an object of class ijtiff::ijtiff\_img). The image to perform the calculation on. To perform this on a file that has not yet been read in, set this argument to the path to that file (a string).

frames\_per\_set The number of frames with which to calculate the successive cross-correlated numbers.

> This may discard some images, for example if 175 frames are in the input and frames\_per\_set = 50, then the last 25 are discarded. If bleaching or/and thresholding are selected, they are performed on the whole image stack before the sectioning is done for calculation of cross-correlated numbers.

overlap

A boolean. If TRUE, the windows used to calculate brightness are overlapped, if FALSE, they are not. For example, for a 20-frame image series with 5 frames per set, if the windows are not overlapped, then the frame sets used are 1-5, 6-10, 11-15 and 16-20; whereas if they are overlapped, the frame sets are 1-5, 2-6, 3-7, 4-8 and so on up to 16-20.

ch1

A natural number. The index of the first channel to use.

ch2

A natural number. The index of the second channel to use.

thresh

Do you want to apply an intensity threshold prior to calculating cross-correlated number (via autothresholdr::mean\_stack\_thresh())? If so, set your thresholding method here. If this is a single value, that same threshold will be applied to both channels. If this is a length-2 vector or list, then these two thresholds will be applied to channels 1 and 2 respectively. A value of NA for either channel gives no thresholding for that channel.

detrend

Detrend your data with detrendr::img\_detrend\_rh(). This is the best known detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. To detrend one channel and not the other, specify this as a length 2 vector.

quick

FALSE repeats the detrending procedure (which has some inherent randomness) a few times to hone in on the best detrend. TRUE is quicker, performing the routine only once. FALSE is better.

filt

Do you want to smooth (filt = 'smooth') or median (filt = 'median') filter the cross-correlated number image using smooth\_filter() or median\_filter() respectively? If selected, these are invoked here with a filter radius of 1 and with the option na\_count = TRUE. A value of NA for either channel gives no thresholding for that channel. If you want to smooth/median filter the cross-correlated number image in a different way, first calculate the cross-correlated numbers without filtering (filt = NULL) using this function and then perform your desired filtering routine on the result.

parallel

Would you like to use multiple cores to speed up this function? If so, set the number of cores here, or to use all available cores, use parallel = TRUE.

#### Value

An array where the *i*th slice is the *i*th cross-correlated number image.

#### See Also

```
number().
```

#### **Examples**

```
img <- ijtiff::read_tif(system.file("extdata", "two_ch.tif",
    package = "nandb"
))
cc_nts <- cc_number_timeseries(img, 10,
    thresh = "Huang",
    filt = "median", parallel = 2
)
ijtiff::display(cc_nts[, , 1, 1])</pre>
```

cc\_number\_timeseries\_folder

Cross-correlated number time-series calculations for every image in a folder.

## Description

Perform cc\_number\_timeseries() calculations on all tif images in a folder and save the resulting images to disk.

```
cc_number_timeseries_folder(
  folder_path = ".",
  frames_per_set,
  overlap = FALSE,
```

```
ch1 = 1,
  ch2 = 2,
  thresh = NULL,
  detrend = FALSE,
  quick = FALSE,
 filt = NULL,
 parallel = FALSE
)
```

#### **Arguments**

folder\_path

The path (relative or absolute) to the folder you wish to process.

frames\_per\_set The number of frames with which to calculate the successive cross-correlated numbers.

> This may discard some images, for example if 175 frames are in the input and frames\_per\_set = 50, then the last 25 are discarded. If bleaching or/and thresholding are selected, they are performed on the whole image stack before the sectioning is done for calculation of cross-correlated numbers.

overlap

A boolean. If TRUE, the windows used to calculate brightness are overlapped, if FALSE, they are not. For example, for a 20-frame image series with 5 frames per set, if the windows are not overlapped, then the frame sets used are 1-5, 6-10, 11-15 and 16-20; whereas if they are overlapped, the frame sets are 1-5, 2-6, 3-7, 4-8 and so on up to 16-20.

ch1

A natural number. The index of the first channel to use.

ch2

A natural number. The index of the second channel to use.

thresh

Do you want to apply an intensity threshold prior to calculating cross-correlated number (via autothresholdr::mean\_stack\_thresh())? If so, set your thresholding method here. If this is a single value, that same threshold will be applied to both channels. If this is a length-2 vector or list, then these two thresholds will be applied to channels 1 and 2 respectively. A value of NA for either channel gives no thresholding for that channel.

detrend

Detrend your data with detrendr::img\_detrend\_rh(). This is the best known detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. To detrend one channel and not the other, specify this as a length 2 vector.

quick

FALSE repeats the detrending procedure (which has some inherent randomness) a few times to hone in on the best detrend. TRUE is quicker, performing the routine only once. FALSE is better.

filt

Do you want to smooth (filt = 'smooth') or median (filt = 'median') filter the cross-correlated number image using smooth\_filter() or median\_filter() respectively? If selected, these are invoked here with a filter radius of 1 and with the option na\_count = TRUE. A value of NA for either channel gives no thresholding for that channel. If you want to smooth/median filter the cross-correlated number image in a different way, first calculate the cross-correlated numbers without filtering (filt = NULL) using this function and then perform your desired filtering routine on the result.

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parallel

Would you like to use multiple cores to speed up this function? If so, set the number of cores here, or to use all available cores, use parallel = TRUE.

## See Also

```
cc_number_timeseries()
```

## **Examples**

```
## Not run:
setwd(tempdir())
ijtiff::write_tif(img, "a.tif")
ijtiff::write_tif(img, "ab.tif")
cc_number_timeseries_folder(frames_per_set = 25)
list.files()
## End(Not run)
```

cross\_var

Calculate the cross-variance of two vectors.

## **Description**

The cross-variance function is defined in the reference.

### Usage

```
cross_var(x, y)
```

#### **Arguments**

x A numeric vector.

y A numeric vector with the same length as x.

#### Value

A number

### References

Digman, MA, Wiseman, PW, Choi, C, Horwitz, AR, Gratton, E (2009). Stoichiometry of molecular complexes at adhesions in living cells. Proc. Natl. Acad. Sci. U.S.A., 106, 7:2170-5.

## **Examples**

```
cross_var(0:3, 2:5)
```

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cross\_var\_pillars

Calculate the cross-variance of corresponding pillars of 3d arrays.

## **Description**

The cross-variance function is defined in the reference.

## Usage

```
cross_var_pillars(x, y)
```

## Arguments

x A 3-dimensional array.

y A 3-dimensional array with the same dimensions as x.

#### **Details**

Pillar i, j of the 3-dimensional array arr is arr[i,j,].

## Value

A matrix.

## **Examples**

```
x <- array(1:27, dim = rep(3, 3))
y <- array(0:26, dim = rep(3, 3))
cross_var_pillars(x, y)</pre>
```

matrix\_raster\_plot

Make a raster plot of a matrix.

## Description

Given a matrix mat, make a raster plot of the matrix whereby in the plot, the pixel at x = i, y = j has colour based on the value of mat[i,j] and the x axis points right and the y axis points down (see 'Details').

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#### Usage

```
matrix_raster_plot(
  mat,
  scale_name = "scale",
  limits = NULL,
  ranges = NULL,
  range_names = NULL,
  colours = NULL,
  na_colour = "black",
  clip = FALSE,
  clip_low = FALSE,
  clip_high = FALSE,
  log_trans = FALSE,
  breaks = NULL,
  include_breaks = NULL
)
```

## **Arguments**

The matrix you wish to plot. mat

scale\_name A string. The title of the color scale on the right of the plot.

This gives the user the option to set all values outside a certain range to their limits nearest value within that range (if clip = TRUE) or to NA (if clip = FALSE. For

example, to set all values outside the range [1.5, 2.6) to NA, use limits = c(1.5, 2.6), clip = FALSE.

The colour range will cover all values within these specified limits.

A numeric vector. If you want specific ranges of values to have the same color, specify these ranges via an increasing numeric vector. For example, if you want the ranges 0.5-1.2 and 1.2-3, use ranges = c(0.5, 1.2, 3). If ranges is specified as a number (a numeric vector of length 1) n, this is equivalent to setting ranges to be n equal-length intervals within the range of the matrix, i.e. it is equivalent to setting 'ranges = seq(min(mat), max(mat), length.out = n

• 1). At most one of rangesandlimits should be set. If ranges is set, the behaviour for values which are not in any of the ranges are set by the cliparguments as in thelimits' argument.

A character vector. If your colour scale is discrete, here you can set the names which will label each range in the legend.

If you have set ranges, here you may specify which colors you wish to colour each range. It must have the same length as the number of intervals you have specified in ranges. If you have not specified ranges, here you may specify the colours (to be passed to ggplot2::scale\_fill\_gradientn()) to create the continuous colour band. It is specified as a character vector, with the colors specified either as the values in colors() or as in the value of the rgb() function. Note that this allows the use of grDevices::rainbow() and friends. The

Which colour should the NA pixels be? Default is black.

default uses viridis::viridis().

ranges

range\_names

colours

na\_colour

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clip	If either limits or ranges are set (one should never set both), there may be values that fall outside the specified limits/ranges. If clip = TRUE, values outside these limits/ranges are set to their nearest values within them, but if clip = FALSE, these values are set to NA. Note that setting clip = TRUE is equivalent to setting both clip_low and clip_high to TRUE.
clip_low	Setting this to TRUE (and leaving clip = FALSE, clip_high = FALSE) will set all values falling below the specified limits/ranges to their nearest value within them, but all values falling above those limits/ranges will be set to NA.
clip_high	Setting this to TRUE (and leaving clip = FALSE, clip_low = FALSE) will set all values falling above the specified limits/ranges to their nearest value within them, but all values falling below those limits/ranges will be set to NA.
log_trans	Do you want to log-transform the colour scaling?
breaks	Where do you want tick marks to appear on the legend colour scale?
include_breaks	If you don't want to specify all the breaks, but you want some specific ones to be included on the legend colour scale, specify those specific ones here.

#### Value

In the graphics console, a raster plot (via ggplot2::geom\_raster()) will appear with the matrix values represented as pixel colours, with a named scale bar.

## **Examples**

```
img <- ijtiff::read_tif(system.file("extdata", "50.tif", package = "nandb"))</pre>
ijtiff::display(img[, , 1, 1])
matrix_raster_plot(img[, , 1, 1])
b <- brightness(img, def = "B", detrend = FALSE, thresh = "Huang")</pre>
matrix_raster_plot(b, scale_name = "brightness")
matrix_raster_plot(b, scale_name = "brightness", log_trans = TRUE)
matrix_raster_plot(b,
  scale_name = "brightness", log_trans = TRUE,
  include_breaks = 1.35
)
matrix_raster_plot(b,
  scale_name = "brightness", log_trans = TRUE,
  breaks = 1:3
)
matrix_raster_plot(b,
  scale_name = "brightness",
  ranges = seq(0.5, 3, length.out = 6),
  range_names = paste0(1:5, "mer")
)
matrix_raster_plot(b,
  scale_name = "brightness",
  ranges = seq(0.5, 3, length.out = 6),
  range_names = paste0(1:5, "mer"), log_trans = TRUE
)
matrix_raster_plot(b,
  scale_name = "brightness",
```

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```
include_breaks = 1.25, range_names = NULL,
  log_trans = FALSE
)
matrix_raster_plot(b,
  scale_name = "brightness",
  include_breaks = 1.25, log_trans = TRUE
)
matrix_raster_plot(b,
  scale_name = "brightness",
  limits = c(1, 1.25), clip = TRUE
)
matrix_raster_plot(b,
  scale_name = "brightness",
  include_breaks = 1.25
)
```

median\_filter

Smooth and median filters with options for handling NAs.

## **Description**

These are alternatives to EBImage::filter2() and EBImage::medianFilter() for smooth and median filtering respectively. These functions have many options for dealing with NA values which EBImage's functions lack.

#### Usage

```
median_filter(mat, size = 1L, na_rm = FALSE, na_count = FALSE)
smooth_filter(mat, size = 1L, na_rm = FALSE, na_count = FALSE)
```

### Arguments

mat A matrix (representing an image).
size An integer; the median filter radius.

na\_rm Should NAs be ignored?

NAs, NA is returned but if the NAs are in the minority, they are ignored as in

median(x,na.rm = TRUE).

#### **Details**

The behavior at image boundaries is such as the source image has been padded with pixels whose values equal the nearest border pixel value.

### Value

A matrix (the median filtered image).

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#### **Examples**

```
m <- matrix(1:9, nrow = 3)
m[2:3, 2:3] <- NA
print(m)
median_filter(m)
median_filter(m, na_rm = TRUE)
median_filter(m, na_count = TRUE)
smooth_filter(m)
smooth_filter(m, na_rm = TRUE)
smooth_filter(m, na_rm = TRUE)</pre>
```

nandb

nandb: Number and brightness in R.

### **Description**

The nandb package gives functions for calculation of molecular number and brightness from images, as detailed in Digman et al. 2008. It comes with an implementation of the novel 'automatic detrending' technique.

## References

Digman MA, Dalal R, Horwitz AF, Gratton E. Mapping the Number of Molecules and Brightness in the Laser Scanning Microscope. Biophysical Journal. 2008;94(6):2320-2332. doi: 10.1529/biophysj.107.114645.

nb-img-classes

Number and brightness image classes.

## Description

The number\_img and brightness\_img classes are designed to hold objects which are images calculated from the *number and brightness* technique.

```
number_img(img, def, thresh, swaps, filt)
brightness_img(img, def, thresh, swaps, filt)
```

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#### **Arguments**

The calculated number or brightness image. img def The number or brightness definition used. thresh A positive integer, possibly an object of class autothresholdr::th. If the different channels of the image had different thresholds, this argument may be specified as a vector or list (of positive integers, possibly objects of class autothresholdr::th), one element for each channel. A non-negative integer with an attribute auto. If the different channels of the imswaps age had different swaps, this argument may be specified as a list (of non-negative integers with attributes auto), one element for each channel. For undetrended images, set swaps = NA. filt A string, the filtering method used. Must be either "mean" or "median", or NA for no filtering. If the different channels of the image had different filters, this may be specified as a character vector, one element for each channel.

#### **Details**

An object of class number\_img or brightness\_img is a 4-dimensional array of real numbers in the mould of an ijtiff\_img (indexed as img[y,x,channel,frame]) with 4 attributes:

def Are we using the "N" or "n" definition of number, or the "B" or "epsilon" definition of brightness?

thresh A positive integer, possibly an object of class autothresholdr::th detailing which threshold and thresholding method was used in preprocessing (in the multi-channel case, one threshold per channel is given).

swaps A non-negative integer indicating the number of swaps Robin Hood detrending, with an attribute auto which is a logical indicating whether or not the parameter was chosen automatically (in the multi-channel case, one threshold per channel is given).

filt Was mean or median filtering used in postprocessing?

#### Value

An object of class number\_img or brightness\_img.

nb-ts-img-classes Number and brightness time series image classes.

#### **Description**

The number\_ts\_img and brightness\_ts\_img classes are designed to hold objects which are images calculated from the *number and brightness* technique.

```
number_ts_img(img, def, frames_per_set, overlapped, thresh, swaps, filt)
brightness_ts_img(img, def, frames_per_set, overlapped, thresh, swaps, filt)
```

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#### **Arguments**

img The calculated number or brightness time series image series.

def The number or brightness definition used.

frames\_per\_set The number of frames used in the calculation of each point in the number or

brightness time series.

overlapped A boolean. TRUE indicates that the windows used to calculate consecutive bright-

nesses over time were overlapped, FALSE indicates that they were not.

thresh A positive integer, possibly an object of class autothresholdr::th. If the different

channels of the image had different thresholds, this argument may be specified as a vector or list (of positive integers, possibly objects of class autothresholdr::th),

one element for each channel.

swaps A non-negative integer with an attribute auto. If the different channels of the im-

age had different swaps, this argument may be specified as a list (of non-negative integers with attributes auto), one element for each channel. For undetrended

images, set swaps = NA.

filt A string, the filtering method used. Must be either "mean" or "median", or NA

for no filtering. If the different channels of the image had different filters, this

may be specified as a character vector, one element for each channel.

#### **Details**

An object of class number\_ts\_img or brightness\_ts\_img is a 3- or 4-dimensional array of real numbers with 4 attributes:

def Are we using the "N" or "n" definition of number, or the "B" or "epsilon" definition of brightness?

thresh A positive integer, possibly an object of class autothresholdr::th detailing which threshold and thresholding method was used in preprocessing (in the multi-channel case, one threshold per channel is given).

swaps A non-negative integer indicating the number of swaps used for Robin Hood detrending, with an attribute auto which is a logical indicating whether or not the parameter was chosen automatically (in the multi-channel case, one swaps per channel is given).

frames\_per\_set A positive integer detailing how many frames were used in the calculation of each point in the number or brightness time series.

overlapped A boolean. TRUE indicates that the windows used to calculate consecutive brightnesses over time were overlapped, FALSE indicates that they were not.

#### Value

An object of class number\_ts\_img or brightness\_ts\_img.

#### See Also

number\_timeseries(), brightness\_timeseries().

number number

number

Calculate number from image series.

## Description

Given a time stack of images, number() performs a calculation of the number for each pixel.

## Usage

```
number(
  img,
  def,
  thresh = NULL,
  detrend = FALSE,
  quick = FALSE,
  filt = NULL,
  s = 1,
  offset = 0,
  readout_noise = 0,
  gamma = 1,
  parallel = FALSE
)
```

## Arguments

img	A 4-dimensional array of images indexed by img[y,x,channel,frame] (an object of class ijtiff::ijtiff_img). The image to perform the calculation on. To perform this on a file that has not yet been read in, set this argument to the path to that file (a string).
def	A character. Which definition of number do you want to use, "n" or "N"?
thresh	The threshold or thresholding method (see autothresholdr::mean_stack_thresh()) to use on the image prior to detrending and number calculations. If there are many channels, this may be specified as a vector or list, one element for each channel.
detrend	Detrend your data with detrendr::img_detrend_rh(). This is the best known detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. If there are many channels, this may be specified as a vector, one element for each channel.
quick	FALSE repeats the detrending procedure (which has some inherent randomness) a few times to hone in on the best detrend. TRUE is quicker, performing the routine only once. FALSE is better.
filt	Do you want to smooth (filt = 'mean') or median (filt = 'median') filter the number image using smooth_filter() or median_filter() respectively? If selected, these are invoked here with a filter radius of 1 (with corners included, so each median is the median of 9 elements) and with the option na_count =

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TRUE. If you want to smooth/median filter the number image in a different way, first calculate the numbers without filtering (filt = NULL) using this function and then perform your desired filtering routine on the result. If there are many channels, this may be specified as a vector, one element for each channel.

s A positive number. The S-factor of microscope acquisition.

offset, readout\_noise

Microscope acquisition parameters. See reference Dalal et al.

gamma

Factor for correction of number n due to the illumination profile. The default (gamma = 1) has no effect. Changing gamma will have the effect of dividing the result by gamma, so the result with gamma = 0.5 is two times the result with gamma = 1. For a Gaussian illumination profile, use gamma = 0.3536; for a Gaussian-Lorentzian illumination profile, use gamma = 0.0760.

parallel

Would you like to use multiple cores to speed up this function? If so, set the number of cores here, or to use all available cores, use parallel = TRUE.

#### Value

A matrix, the number image.

#### References

Digman MA, Dalal R, Horwitz AF, Gratton E. Mapping the Number of Molecules and Brightness in the Laser Scanning Microscope. Biophysical Journal. 2008;94(6):2320-2332. doi: 10.1529/biophysj.107.114645.

Dalal, RB, Digman, MA, Horwitz, AF, Vetri, V, Gratton, E (2008). Determination of particle number and brightness using a laser scanning confocal microscope operating in the analog mode. Microsc. Res. Tech., 71, 1:69-81. doi: 10.1002/jemt.20526.

Hur K-H, Macdonald PJ, Berk S, Angert CI, Chen Y, Mueller JD (2014) Quantitative Measurement of Brightness from Living Cells in the Presence of Photodepletion. PLoS ONE 9(5): e97440. doi: 10.1371/journal.pone.0097440.

## **Examples**

```
img <- ijtiff::read_tif(system.file("extdata", "50.tif", package = "nandb"))
ijtiff::display(img[, , 1, 1])
num <- number(img, "N", thresh = "Huang")
num <- number(img, "n", thresh = "tri")</pre>
```

number\_folder

Number calculations for every image in a folder.

#### **Description**

Perform number() calculations on all tif images in a folder and save the resulting number images to disk.

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#### Usage

```
number_folder(
  folder_path = ".",
  def,
  thresh = NULL,
  detrend = FALSE,
  quick = FALSE,
  filt = NULL,
  s = 1,
  offset = 0,
  readout_noise = 0,
  gamma = 1,
  parallel = FALSE
)
```

## **Arguments**

folder\_path The path (relative or absolute) to the folder you wish to process.

def A character. Which definition of number do you want to use, "n" or "N"?

thresh The threshold or thresholding method (see autothresholdr::mean\_stack\_thresh())

to use on the image prior to detrending and number calculations. If there are many channels, this may be specified as a vector or list, one element for each

channel.

detrend Detrend your data with detrendr::img\_detrend\_rh(). This is the best known

detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. If there are many channels, this

may be specified as a vector, one element for each channel.

quick FALSE repeats the detrending procedure (which has some inherent randomness)

a few times to hone in on the best detrend. TRUE is quicker, performing the

routine only once. FALSE is better.

filt Do you want to smooth (filt = 'mean') or median (filt = 'median') filter the

number image using smooth\_filter() or median\_filter() respectively? If selected, these are invoked here with a filter radius of 1 (with corners included, so each median is the median of 9 elements) and with the option na\_count = TRUE. If you want to smooth/median filter the number image in a different way, first calculate the numbers without filtering (filt = NULL) using this function and then perform your desired filtering routine on the result. If there are many

channels, this may be specified as a vector, one element for each channel.

s A positive number. The S-factor of microscope acquisition.

offset Microscope acquisition parameters. See reference Dalal et al.

readout\_noise Microscope acquisition parameters. See reference Dalal et al.

gamma Factor for correction of number n due to the illumination profile. The default

(gamma = 1) has no effect. Changing gamma will have the effect of dividing the result by gamma, so the result with gamma = 0.5 is two times the result with gamma = 1. For a Gaussian illumination profile, use gamma = 0.3536; for

a Gaussian-Lorentzian illumination profile, use gamma = 0.0760.

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parallel

Would you like to use multiple cores to speed up this function? If so, set the number of cores here, or to use all available cores, use parallel = TRUE.

## Note

Extreme number values (of magnitude greater than 3.40282e+38) will be written to the TIFF file as NA, since TIFF files cannot handle such huge numbers.

#### See Also

```
number()
```

## **Examples**

```
## Not run:
setwd(tempdir())
img <- ijtiff::read_tif(system.file("extdata", "50.tif", package = "nandb"))
ijtiff::write_tif(img, "img2.tif")
number_folder(def = "n", thresh = "Huang", parallel = 2)
## End(Not run)</pre>
```

number\_timeseries

Create a number time-series.

#### **Description**

Given a stack of images img, use the first frames\_per\_set of them to create one number image, the next frames\_per\_set of them to create the next number image and so on to get a time-series of number images.

```
number_timeseries(
   img,
   def,
   frames_per_set,
   overlap = FALSE,
   thresh = NULL,
   detrend = FALSE,
   quick = FALSE,
   filt = NULL,
   s = 1,
   offset = 0,
   readout_noise = 0,
   gamma = 1,
   parallel = FALSE
)
```

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#### **Arguments**

img A 4-dimensional array of images indexed by img[y,x,channel,frame] (an ob-

ject of class ijtiff::ijtiff\_img). The image to perform the calculation on. To perform this on a file that has not yet been read in, set this argument to the path to

that file (a string).

def A character. Which definition of number do you want to use, "n" or "N"?

frames\_per\_set The number of frames with which to calculate the successive numbers.

overlap A boolean. If TRUE, the windows used to calculate brightness are overlapped, if

FALSE, they are not. For example, for a 20-frame image series with 5 frames per set, if the windows are not overlapped, then the frame sets used are 1-5, 6-10, 11-15 and 16-20; whereas if they are overlapped, the frame sets are 1-5, 2-6,

3-7, 4-8 and so on up to 16-20.

thresh The threshold or thresholding method (see autothresholdr::mean\_stack\_thresh())

to use on the image prior to detrending and number calculations. If there are many channels, this may be specified as a vector or list, one element for each

channel.

detrend Detrend your data with detrendr::img\_detrend\_rh(). This is the best known

detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. If there are many channels, this

may be specified as a vector, one element for each channel.

quick FALSE repeats the detrending procedure (which has some inherent randomness)

a few times to hone in on the best detrend. TRUE is quicker, performing the

routine only once. FALSE is better.

filt Do you want to smooth (filt = 'mean') or median (filt = 'median') filter the

number image using smooth\_filter() or median\_filter() respectively? If selected, these are invoked here with a filter radius of 1 (with corners included, so each median is the median of 9 elements) and with the option na\_count = TRUE. If you want to smooth/median filter the number image in a different way, first calculate the numbers without filtering (filt = NULL) using this function and then perform your desired filtering routine on the result. If there are many

channels, this may be specified as a vector, one element for each channel.

s A positive number. The S-factor of microscope acquisition.

offset Microscope acquisition parameters. See reference Dalal et al.

readout\_noise Microscope acquisition parameters. See reference Dalal et al.

gamma Factor for correction of number n due to the illumination profile. The default

(gamma = 1) has no effect. Changing gamma will have the effect of dividing the result by gamma, so the result with gamma = 0.5 is two times the result with gamma = 1. For a Gaussian illumination profile, use gamma = 0.3536; for

a Gaussian-Lorentzian illumination profile, use gamma = 0.0760.

parallel Would you like to use multiple cores to speed up this function? If so, set the

number of cores here, or to use all available cores, use parallel = TRUE.

#### **Details**

This may discard some images, for example if 175 frames are in the input and frames\_per\_set = 50, then the last 25 are discarded. If detrending is selected, it is performed on the whole image stack before the sectioning is done for calculation of numbers.

## Value

An object of class number\_ts\_img.

#### See Also

```
number().
```

#### **Examples**

```
img <- ijtiff::read_tif(system.file("extdata", "50.tif", package = "nandb"))
nts <- number_timeseries(img, "n", frames_per_set = 20, thresh = "Huang")</pre>
```

```
number_timeseries_folder
```

Number time-series calculations for every image in a folder.

## Description

Perform number\_timeseries() calculations on all tif images in a folder and save the resulting number images to disk.

```
number_timeseries_folder(
  folder_path = ".",
  def,
  frames_per_set,
  overlap = FALSE,
  thresh = NULL,
  detrend = FALSE,
  quick = FALSE,
  filt = NULL,
  s = 1,
  offset = 0,
  readout_noise = 0,
  gamma = 1,
  parallel = FALSE
)
```

#### **Arguments**

folder\_path The path (relative or absolute) to the folder you wish to process.

def A character. Which definition of number do you want to use, "n" or "N"?

frames\_per\_set The number of frames with which to calculate the successive numbers.

overlap A boolean. If TRUE, the windows used to calculate brightness are overlapped, if

FALSE, they are not. For example, for a 20-frame image series with 5 frames per set, if the windows are not overlapped, then the frame sets used are 1-5, 6-10, 11-15 and 16-20; whereas if they are overlapped, the frame sets are 1-5, 2-6,

3-7, 4-8 and so on up to 16-20.

thresh The threshold or thresholding method (see autothresholdr::mean\_stack\_thresh())

to use on the image prior to detrending and number calculations. If there are many channels, this may be specified as a vector or list, one element for each

channel.

detrend Detrend your data with detrendr::img\_detrend\_rh(). This is the best known

detrending method for brightness analysis. For more fine-grained control over your detrending, use the detrendr package. If there are many channels, this

may be specified as a vector, one element for each channel.

quick FALSE repeats the detrending procedure (which has some inherent randomness)

a few times to hone in on the best detrend. TRUE is quicker, performing the

routine only once. FALSE is better.

filt Do you want to smooth (filt = 'mean') or median (filt = 'median') filter the

number image using smooth\_filter() or median\_filter() respectively? If selected, these are invoked here with a filter radius of 1 (with corners included, so each median is the median of 9 elements) and with the option na\_count = TRUE. If you want to smooth/median filter the number image in a different way, first calculate the numbers without filtering (filt = NULL) using this function and then perform your desired filtering routine on the result. If there are many channels, this may be specified as a vector, one element for each channel.

s A positive number. The S-factor of microscope acquisition.

offset Microscope acquisition parameters. See reference Dalal et al.

readout\_noise Microscope acquisition parameters. See reference Dalal et al.

gamma Factor for correction of number n due to the illumination profile. The default

(gamma = 1) has no effect. Changing gamma will have the effect of dividing the result by gamma, so the result with gamma = 0.5 is two times the result with gamma = 1. For a Gaussian illumination profile, use gamma = 0.3536; for

a Gaussian-Lorentzian illumination profile, use gamma = 0.0760.

parallel Would you like to use multiple cores to speed up this function? If so, set the

number of cores here, or to use all available cores, use parallel = TRUE.

#### Note

Extreme number values (of magnitude greater than 3.40282e+38) will be written to the TIFF file as NA, since TIFF files cannot handle such huge numbers.

## See Also

```
number_timeseries()
```

## Examples

```
## Not run:
setwd(tempdir())
img <- ijtiff::read_tif(system.file("extdata", "50.tif", package = "nandb"))
ijtiff::write_tif(img, "img1.tif")
ijtiff::write_tif(img, "img2.tif")
number_timeseries_folder(def = "n", thresh = "Huang", frames_per_set = 20)
## End(Not run)</pre>
```

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