

Package ‘tabula’

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Title Analysis, Seriation and Visualization of Archaeological Count Data

Version 1.6.1

Maintainer Nicolas Frerebeau <nicolas.frerebeau@u-bordeaux-montaigne.fr>

Description An easy way to examine archaeological count data. This package provides a convenient and reproducible toolkit for relative dating by matrix seriation (reciprocal ranking, CA-based seriation). It also provides several tests and measures of diversity: heterogeneity and evenness (Brillouin, Shannon, Simpson, etc.), richness and rarefaction (Chao1, Chao2, ACE, ICE, etc.), turnover and similarity (Brainerd-Robinson, etc.). The package make it easy to visualize count data and statistical thresholds: rank vs abundance plots, heatmaps, Ford (1962) and Bertin (1977) diagrams.

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URL <https://packages.tesselle.org/tabula/>,
<https://github.com/tesselle/tabula>

BugReports <https://github.com/tesselle/tabula/issues>

Depends R (>= 3.3)

Imports arkhe (>= 0.3.0), dimensio, ggplot2, grDevices, methods, rlang, stats, utils

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Collate 'AllClasses.R' 'AllGenerics.R' 'coerce.R' 'deprecate.R'
'ggplot2.R' 'index_diversity.R' 'index_heterogeneity.R'
'index_rarefaction.R' 'index_richness.R' 'index_similarity.R'
'index_turnover.R' 'mutators.R' 'permute.R' 'plot_bertin.R'
'plot_diversity.R' 'plot_ford.R' 'plot_heatmap.R' 'plot_rank.R'

```
'plot_spot.R' 'reexport.R' 'refine_ca.R' 'seriate_average.R'
'seriate_rank.R' 'show.R' 'statistics.R' 'subset.R'
'tabula-package.R' 'test_diversity.R' 'utilities.R'
'validate.R' 'zzz.R'
```

NeedsCompilation no

Author Nicolas Frerebeau [aut, cre] (<<https://orcid.org/0000-0001-5759-4944>>),
 Brice Lebrun [ctb] (<<https://orcid.org/0000-0001-7503-8685>>),
 Matthew Peebles [ctb] (<<https://orcid.org/0000-0003-4496-623X>>),
 Ben Marwick [ctb] (<<https://orcid.org/0000-0001-7879-4531>>),
 Anne Philippe [ctb] (<<https://orcid.org/0000-0002-5331-5087>>),
 Jean-Baptiste Fourvel [ctb] (<<https://orcid.org/0000-0002-1061-4642>>)

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R topics documented:

DiversityIndex	2
heterogeneity-index	3
independance	8
mutator	9
PermutationOrder-class	10
plot_bar	11
plot_diversity	13
plot_line	14
plot_matrix	15
plot_spot	16
RefineCA-class	17
richness-index	18
seriation	22
similarity	26
subset	28
test_diversity	29
turnover-index	30

Index	33
--------------	-----------

Description

An S4 class to represent a diversity measure.

Slots

names A `character` vector giving the sample names.
values A `numeric` vector giving the diversity index values.
size A `integer` vector giving the sample sizes.
simulation A four columns `numeric` matrix giving the diversity measures for the simulated assemblage (sample size, mean estimate, lower and upper boundaries of the confidence interval).
method A `character` string specifying the method used.

Subset

In the code snippets below, `x` is a `DiversityIndex` object.

`x[[i]]` Extracts information from a slot selected by subscript `i`. `i` is a length-one `character` vector. Returns the corresponding slot values.

Author(s)

N. Frerebeau

See Also

Other class: `PermutationOrder-class`, `RefineCA-class`

heterogeneity-index *Heterogeneity and Evenness*

Description

- `index_heterogeneity()` returns an heterogeneity or dominance index.
- `index_evenness()` returns an evenness measure.
- `bootstrap_*`() and `jackknife_*`() perform bootstrap/jackknife resampling.

Usage

```
index_heterogeneity(object, ...)

simulate_heterogeneity(object, ...)

bootstrap_heterogeneity(object, ...)

jackknife_heterogeneity(object, ...)

index_evenness(object, ...)

simulate_evenness(object, ...)
```

```
bootstrap_evenness(object, ...)

jackknife_evenness(object, ...)

## S4 method for signature 'CountMatrix'
index_heterogeneity(
  object,
  method = c("berger", "brillouin", "mcintosh", "shannon", "simpson"),
  ...
)

## S4 method for signature 'CountMatrix'
simulate_heterogeneity(
  object,
  method = c("berger", "brillouin", "mcintosh", "shannon", "simpson"),
  quantiles = TRUE,
  level = 0.8,
  step = 1,
  n = 1000,
  progress = getOption("tabula.progress"),
  ...
)

## S4 method for signature 'CountMatrix'
bootstrap_heterogeneity(
  object,
  method = c("berger", "brillouin", "mcintosh", "shannon", "simpson"),
  probs = c(0.05, 0.95),
  n = 1000,
  ...
)

## S4 method for signature 'CountMatrix'
jackknife_heterogeneity(
  object,
  method = c("berger", "brillouin", "mcintosh", "shannon", "simpson"),
  ...
)

## S4 method for signature 'CountMatrix'
index_evenness(
  object,
  method = c("shannon", "brillouin", "mcintosh", "simpson"),
  ...
)

## S4 method for signature 'CountMatrix'
```

```

simulate_evenness(
  object,
  method = c("shannon", "brillouin", "mcintosh", "simpson"),
  quantiles = TRUE,
  level = 0.8,
  step = 1,
  n = 1000,
  progress = getOption("tabula.progress"),
  ...
)

## S4 method for signature 'CountMatrix'
bootstrap_evenness(
  object,
  method = c("berger", "brillouin", "mcintosh", "shannon", "simpson"),
  probs = c(0.05, 0.95),
  n = 1000,
  ...
)

## S4 method for signature 'CountMatrix'
jackknife_evenness(
  object,
  method = c("berger", "brillouin", "mcintosh", "shannon", "simpson"),
  ...
)

```

Arguments

object	A $m \times p$ matrix of count data (typically a <code>CountMatrix</code> object).
...	Further arguments to be passed to internal methods.
method	A <code>character</code> string specifying the index to be computed (see details). Any unambiguous substring can be given.
quantiles	A <code>logical</code> scalar: should sample quantiles be used as confidence interval? If <code>TRUE</code> (the default), sample quantiles are used as described in Kintigh (1989), else quantiles of the normal distribution are used.
level	A length-one <code>numeric</code> vector giving the confidence level.
step	A non-negative <code>integer</code> giving the increment of the sample size. Only used if <code>simulate</code> is <code>TRUE</code> .
n	A non-negative <code>integer</code> giving the number of bootstrap replications.
progress	A <code>logical</code> scalar: should a progress bar be displayed?
probs	A <code>numeric</code> vector of probabilities with values in $[0, 1]$ (see <code>stats:::quantile()</code>).

Details

Diversity measurement assumes that all individuals in a specific taxa are equivalent and that all types are equally different from each other (Peet 1974). A measure of diversity can be achieved by

using indices built on the relative abundance of taxa. These indices (sometimes referred to as non-parametric indices) benefit from not making assumptions about the underlying distribution of taxa abundance: they only take relative abundances of the species that are present and species richness into account. Peet (1974) refers to them as indices of *heterogeneity*.

Diversity indices focus on one aspect of the taxa abundance and emphasize either *richness* (weighting towards uncommon taxa) or dominance (weighting towards abundant taxa; Magurran 1988).

Evenness is a measure of how evenly individuals are distributed across the sample.

The following heterogeneity index and corresponding evenness measures are available (see Magurran 1988 for details):

`berger` Berger-Parker dominance index. The Berger-Parker index expresses the proportional importance of the most abundant type. This metric is highly biased by sample size and richness, moreover it does not make use of all the information available from sample.

`brillouin` Brillouin diversity index. The Brillouin index describes a known collection: it does not assume random sampling in an infinite population. Pielou (1975) and Laxton (1978) argues for the use of the Brillouin index in all circumstances, especially in preference to the Shannon index.

`mcintosh` McIntosh dominance index. The McIntosh index expresses the heterogeneity of a sample in geometric terms. It describes the sample as a point of a S -dimensional hypervolume and uses the Euclidean distance of this point from the origin.

`shannon` Shannon-Wiener diversity index. The Shannon index assumes that individuals are randomly sampled from an infinite population and that all taxa are represented in the sample (it does not reflect the sample size). The main source of error arises from the failure to include all taxa in the sample: this error increases as the proportion of species discovered in the sample declines (Peet 1974, Magurran 1988). The maximum likelihood estimator (MLE) is used for the relative abundance, this is known to be negatively biased by sample size.

`simpson` Simpson dominance index for finite sample. The Simpson index expresses the probability that two individuals randomly picked from a finite sample belong to two different types. It can be interpreted as the weighted mean of the proportional abundances. This metric is a true probability value, it ranges from 0 (perfectly uneven) to 1 (perfectly even).

The `berger`, `mcintosh` and `simpson` methods return a *dominance* index, not the reciprocal or inverse form usually adopted, so that an increase in the value of the index accompanies a decrease in diversity.

Value

- `index_heterogeneity()`, `index_evenness()` and `simulate_evenness()` return a [DiversityIndex](#) object.
- `bootstrap_*`() and `jackknife_*`() return a [data.frame](#).

Note

Ramanujan approximation is used for $x!$ computation if $x > 170$.

Author(s)

N. Frerebeau

References

- Berger, W. H. & Parker, F. L. (1970). Diversity of Planktonic Foraminifera in Deep-Sea Sediments. *Science*, 168(3937), 1345-1347. doi: [10.1126/science.168.3937.1345](https://doi.org/10.1126/science.168.3937.1345).
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- Simpson, E. H. (1949). Measurement of Diversity. *Nature*, 163(4148), 688-688. doi: [10.1038/163688a0](https://doi.org/10.1038/163688a0).

See Also

[plot_diversity\(\)](#), [similarity\(\)](#), [turnover\(\)](#)
 Other diversity: [richness-index](#), [similarity\(\)](#), [turnover-index](#)

Examples

```
## Coerce dataset to a count matrix
data("chevelon", package = "folio")
chevelon <- as_count(chevelon)

## Shannon diversity index
(index_h <- index_heterogeneity(chevelon, method = "shannon"))
(index_e <- index_evenness(chevelon, method = "shannon"))

## Bootstrap resampling
(boot_h <- bootstrap_heterogeneity(chevelon, method = "shannon"))

## Jackknife resampling
(jack_h <- jackknife_heterogeneity(chevelon, method = "shannon"))
```

independance	<i>Independance</i>
--------------	---------------------

Description

`Independance`

Usage

```
eppm(object, ...)
pvi(object, ...)

## S4 method for signature 'CountMatrix'
eppm(object)

## S4 method for signature 'CountMatrix'
pvi(object)
```

Arguments

object	A CountMatrix object.
...	Currently not used.

Details

Computes for each cell of a numeric matrix one of the following statistic.

Value

A [numeric matrix](#).

EPPM

This positive difference from the column mean percentage (in french "écart positif au pourcentage moyen", EPPM) represents a deviation from the situation of statistical independence. As independence can be interpreted as the absence of relationships between types and the chronological order of the assemblages, EPPM is a useful graphical tool to explore significance of relationship between rows and columns related to seriation (Desachy 2004).

PVI

PVI is calculated for each cell as the percentage to the column theoretical independence value: PVI greater than 1 represent positive deviations from the independence, whereas PVI smaller than 1 represent negative deviations (Desachy 2004).

The PVI matrix allows to explore deviations from independence (an intuitive graphical approach to χ^2), in such a way that a high-contrast matrix has quite significant deviations, with a low risk of being due to randomness (Desachy 2004).

Author(s)

N. Frerebeau

References

Desachy, B. (2004). Le sériographe EPPM: un outil informatisé de sériation graphique pour tableaux de comptages. *Revue archéologique de Picardie*, 3(1), 39-56. doi: [10.3406/pica.2004.2396](https://doi.org/10.3406/pica.2004.2396).

See Also

[plot_ford\(\)](#), [plot_heatmap\(\)](#), [seriate_rank\(\)](#)

Other statistics: [test_diversity\(\)](#)

Examples

```
## Abundance data
## Coerce dataset to a count matrix (data from Desachy 2004)
data("compiegne", package = "folio")
counts <- as_count(compiegne)

## Compute EPPM
counts_eppm <- eppm(counts)

## Compute PVI
counts_pvi <- pvi(counts)
plot_heatmap(counts_eppm)
```

mutator

Get or Set Parts of an Object

Description

Getters and setters to extract or replace parts of an object.

Usage

```
get_index(x)

get_method(x)

get_order(x)

## S4 method for signature 'HeterogeneityIndex'
get_index(x)

## S4 method for signature 'EvennessIndex'
get_index(x)
```

```
## S4 method for signature 'RichnessIndex'
get_index(x)

## S4 method for signature 'DiversityIndex'
get_method(x)
```

Arguments

x An object from which to get or set element(s).

Value

An object of the same sort as `object` with the new values assigned.

Author(s)

N. Frerebeau

See Also

Other mutator: [subset\(\)](#)

PermutationOrder-class

Permutation Order

Description

An S4 class to represent a permutation order.

Slots

`rows` An [integer](#) vector giving the rows permutation.

`columns` An [integer](#) vector giving the columns permutation.

`method` A [character](#) string indicating the seriation method used.

Subset

In the code snippets below, `x` is a `PermutationOrder` object.

`x[[i]]` Extracts information from a slot selected by subscript `i`. `i` is a length-one [character](#) vector. Returns the corresponding slot values.

Author(s)

N. Frerebeau

See Also

Other class: [DiversityIndex](#), [RefineCA-class](#)

plot_bar*Bar Plot*

Description

Plots a Bertin, Ford (battleship curve) or Dice-Leraas diagram.

Usage

```
plot_bertin(object, ...)
plot_ford(object, ...)

## S4 method for signature 'matrix'
plot_bertin(object, threshold = NULL, scale = NULL)

## S4 method for signature 'matrix'
plot_ford(object)

## S4 method for signature 'CountMatrix'
plot_ford(object, EPPM = FALSE)
```

Arguments

object	An abundance matrix to be plotted.
...	Currently not used.
threshold	A function that takes a numeric vector as argument and returns a numeric threshold value (see below). If NULL (the default), no threshold is computed.
scale	A function used to scale each variable, that takes a numeric vector as argument and returns a numeric vector. If NULL (the default), no scaling is performed.
EPPM	A logical scalar: should the EPPM be drawn (see below)?

Details

If EPPM is TRUE and if a relative abundance is greater than the mean percentage of the type, the exceeding part is highlighted.

Value

A [ggplot2::ggplot](#) object.

Bertin Matrix

As de Falguerolles *et al.* (1997) points out: "In abstract terms, a Bertin matrix is a matrix of displays. ... To fix ideas, think of a data matrix, variable by case, with real valued variables. For each variable, draw a bar chart of variable value by case. High-light all bars representing a value above some sample threshold for that variable."

EPPM

This positive difference from the column mean percentage (in french "écart positif au pourcentage moyen", EPPM) represents a deviation from the situation of statistical independence. As independence can be interpreted as the absence of relationships between types and the chronological order of the assemblages, EPPM is a useful graphical tool to explore significance of relationship between rows and columns related to seriation (Desachy 2004).

Author(s)

N. Frerebeau

References

- Bertin, J. (1977). *La graphique et le traitement graphique de l'information*. Paris: Flammarion. Nouvelle Bibliothèque Scientifique.
- de Falguerolles, A., Friedrich, F. & Sawitzki, G. (1997). A Tribute to J. Bertin's Graphical Data Analysis. In W. Badilla & F. Faulbaum (eds.), *SoftStat '97: Advances in Statistical Software 6*. Stuttgart: Lucius & Lucius, p. 11-20.
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- Ford, J. A. (1962). *A quantitative method for deriving cultural chronology*. Washington, DC: Pan American Union. Technical manual 1.

See Also

[eppm\(\)](#)

Other plot: [plot_diversity](#), [plot_line](#), [plot_matrix](#), [plot_spot\(\)](#)

Examples

```
## Abundance data
## Coerce dataset to a count matrix
data("mississippi", package = "folio")
counts1 <- as_count(mississippi)

## Plot a Bertin diagram...
## ...without threshold
plot_bertin(counts1)
## ...with variables scaled to 0-1 and the variable mean as threshold
scale_01 <- function(x) (x - min(x)) / (max(x) - min(x))
plot_bertin(counts1, threshold = mean, scale = scale_01)

## Abundance data
## Coerce dataset to a count matrix (data from Desachy 2004)
data("compiegne", package = "folio")
counts2 <- as_count(compiegne)

## Plot a Ford diagram...
```

```
## ...without threshold  
plot_ford(counts2)  
## ...with EPPM  
plot_ford(counts2, EPPM = TRUE)
```

plot_diversity *Diversity Plot*

Description

Diversity Plot

Usage

```
## S4 method for signature 'DiversityIndex,missing'  
plot(x)
```

Arguments

x A [DiversityIndex](#) object to be plotted.

Author(s)

N. Frerebeau

See Also

[index_heterogeneity\(\)](#), [index_evenness\(\)](#), [index_richness\(\)](#)

Other plot: [plot_bar](#), [plot_line](#), [plot_matrix](#), [plot_spot\(\)](#)

Examples

```
## Coerce data to a count matrix  
data("chevelon", package = "folio")  
chevelon <- as_count(chevelon)  
  
## Assemblage diversity size comparison  
## Warning: this may take a few seconds!  
sim_evenness <- simulate_evenness(chevelon, method = "shannon")  
plot(sim_evenness)  
  
sim_richness <- simulate_richness(chevelon, method = "none")  
plot(sim_richness)
```

plot_line*Line Plot***Description**

Plots a rank *vs* relative abundance diagram.

Usage

```
plot_rank(object, ...)
## S4 method for signature 'matrix'
plot_rank(object, log = NULL, facet = FALSE)
```

Arguments

<code>object</code>	An abundance matrix to be plotted.
<code>...</code>	Further arguments to be passed to internal methods.
<code>log</code>	A <code>character</code> string which contains "x" if the x axis is to be logarithmic, "y" if the y axis is to be logarithmic and "xy" or "yx" if both axes are to be logarithmic (base 10).
<code>facet</code>	A <code>logical</code> scalar: should a matrix of panels defined by case/sample be drawn?

Value

A `ggplot2::ggplot` object.

Author(s)

N. Frerebeau

References

Magurran, A. E. (1988). *Ecological Diversity and its Measurement*. Princeton, NJ: Princeton University Press. doi: [10.1007/9789401573580](https://doi.org/10.1007/9789401573580).

See Also

Other plot: `plot_bar`, `plot_diversity`, `plot_matrix`, `plot_spot()`

Examples

```
## Abundance matrix
## Coerce datasets to a count matrix (data from Desachy 2004)
data("compiegne", package = "folio")
counts <- as_count(compiegne)

## Plot rank vs abundance
```

```
plot_rank(counts)
plot_rank(counts, facet = TRUE)
```

plot_matrix	<i>Heatmap</i>
-------------	----------------

Description

Plots a heatmap.

Usage

```
plot_heatmap(object, ...)
## S4 method for signature 'matrix'
plot_heatmap(object)
```

Arguments

object	An object to be plotted.
...	Further arguments to be passed to internal methods.

Value

A `ggplot2::ggplot` object.

Author(s)

N. Frerebeau

References

Desachy, B. (2004). Le sériographe EPPM: un outil informatisé de sériation graphique pour tableaux de comptages. *Revue archéologique de Picardie*, 3(1), 39-56. doi: [10.3406/pica.2004.2396](https://doi.org/10.3406/pica.2004.2396).

See Also

[pvi\(\)](#)

Other plot: [plot_bar](#), [plot_diversity](#), [plot_line](#), [plot_spot\(\)](#)

Examples

```
## Abundance data (data from Desachy 2004)
data("compiegne", package = "folio")
## Coerce dataset to absolute frequencies
counts <- as_count(compiegne)
## Coerce dataset to relative frequencies
freq <- as_composition(compiegne)

## Plot matrix diagram...
plot_heatmap(counts)
plot_heatmap(freq)

## Presence/absence data
inc <- sample(0:1, size = 100, replace = TRUE)
bin <- IncidenceMatrix(data = inc, nrow = 10, ncol = 10)

plot_heatmap(bin) +
  ggplot2::scale_fill_manual(values = c("TRUE" = "black", "FALSE" = "white"))
```

plot_spot

Spot Plot

Description

Plots a spot matrix.

Usage

```
plot_spot(object, ...)

## S4 method for signature 'matrix'
plot_spot(object, threshold = NULL, diag = TRUE, upper = TRUE, ...)

## S4 method for signature 'dist'
plot_spot(object, diag = FALSE, upper = FALSE, ...)

## S4 method for signature 'OccurrenceMatrix'
plot_spot(object, diag = FALSE, upper = FALSE, ...)
```

Arguments

object	An abundance matrix to be plotted.
...	Extra parameters to be passed to <code>threshold</code> .
threshold	A <code>function</code> that takes a numeric vector as argument and returns a numeric threshold value. If <code>NULL</code> (the default), no threshold is computed.
diag	A <code>logical</code> scalar indicating whether the diagonal of the matrix should be plotted.
upper	A <code>logical</code> scalar indicating whether the upper triangle of the matrix should be plotted.

Details

The spot matrix can be considered as a variant of the [Bertin diagram](#) where the data are first transformed to relative frequencies.

Value

A `ggplot2::ggplot` object.

Note

Adapted from Dan Gopstein's original [idea](#).

Author(s)

N. Frerebeau

See Also

Other plot: `plot_bar`, `plot_diversity`, `plot_line`, `plot_matrix`

Examples

```
## Plot spot diagram of count data...
data("mississippi", package = "folio")
counts <- as_count(mississippi)

### ...without threshold
plot_spot(counts)
### ...with the column means as threshold
plot_spot(counts, threshold = mean)
### ...with the column medians as threshold
plot_spot(counts, threshold = median)
```

Description

An S4 class to store partial bootstrap correspondence analysis results.

Slots

`row_chull` A three columns `numeric` matrix giving the vertices coordinates (x, y) of the samples convex hull and a identifier (`id`) to link each row to a sample.
`row_lengths` A named `numeric` vector giving the convex hull maximum dimension length of samples.
`row_keep` An `integer` vector giving the subscript of the samples to be kept.

`column_chull` A three columns `numeric` matrix giving the vertices coordinates (x, y) of the variables convex hull and a identifier (id) to link each row to a variable.

`column_lengths` A `numeric` vector giving the convex hull maximum dimension length of variables.

`column_keep` An `integer` vector giving the subscript of the variables to be kept.

`cutoff` A length-two `numeric` vector giving the cutoff value for samples and variables selection, respectively.

Subset

In the code snippets below, `x` is a `RefineCA` object.

`x[[i]]` Extracts information from a slot selected by subscript `i`. `i` is a length-one `character` vector. Returns the corresponding slot values.

Author(s)

N. Frerebeau

See Also

`dimensio::CA`

Other class: `DiversityIndex`, `PermutationOrder-class`

richness-index

Richness and Rarefaction

Description

- `index_richness()` returns sample richness. `index_composition()` returns asymptotic species richness.
- `rarefaction()` returns Hurlbert's unbiased estimate of Sander's rarefaction.
- `bootstrap_*`() and `jackknife_*`() perform bootstrap/jackknife resampling.

Usage

```
index_richness(object, ...)
simulate_richness(object, ...)
bootstrap_richness(object, ...)
jackknife_richness(object, ...)
index_composition(object, ...)
```

```
rarefaction(object, ...)

## S4 method for signature 'CountMatrix'
rarefaction(object, sample, method = c("hurlbert"), simplify = TRUE, ...)

## S4 method for signature 'CountMatrix'
index_richness(object, method = c("none", "margalef", "menhinick"), ...)

## S4 method for signature 'CountMatrix'
simulate_richness(
  object,
  method = c("none", "margalef", "menhinick"),
  quantiles = TRUE,
  level = 0.8,
  step = 1,
  n = 1000,
  progress = getOption("tabula.progress"),
  ...
)

## S4 method for signature 'CountMatrix'
bootstrap_richness(
  object,
  method = c("none", "margalef", "menhinick"),
  probs = c(0.05, 0.95),
  n = 1000,
  ...
)

## S4 method for signature 'CountMatrix'
jackknife_richness(object, method = c("none", "margalef", "menhinick"), ...)

## S4 method for signature 'CountMatrix'
index_composition(
  object,
  method = c("chao1", "ace"),
  unbiased = FALSE,
  improved = FALSE,
  k = 10
)

## S4 method for signature 'IncidenceMatrix'
index_composition(
  object,
  method = c("chao2", "ice"),
  unbiased = FALSE,
  improved = FALSE,
  k = 10
```

)

Arguments

object	A $m \times p$ matrix of count data.
...	Further arguments to be passed to internal methods.
sample	A length-one numeric vector giving the sub-sample size.
method	A character string or vector of strings specifying the index to be computed (see details). Any unambiguous substring can be given.
simplify	A logical scalar: should the result be simplified to a matrix? The default value, FALSE, returns a list.
quantiles	A logical scalar: should sample quantiles be used as confidence interval? If TRUE (the default), sample quantiles are used as described in Kintigh (1989), else quantiles of the normal distribution are used.
level	A length-one numeric vector giving the confidence level.
step	A non-negative integer giving the increment of the sample size. Only used if simulate is TRUE.
n	A non-negative integer giving the number of bootstrap replications.
progress	A logical scalar: should a progress bar be displayed?
probs	A numeric vector of probabilities with values in [0, 1] (see stats:::quantile()).
unbiased	A logical scalar. Should the bias-corrected estimator be used? Only used with "chao1" or "chao2" (improved) estimator.
improved	A logical scalar. Should the improved estimator be used? Only used with "chao1" or "chao2".
k	A length-one numeric vector giving the threshold between rare/infrequent and abundant/frequent species. Only used if method is "ace" or "ice".

Details

The number of different taxa, provides an instantly comprehensible expression of diversity. While the number of taxa within a sample is easy to ascertain, as a term, it makes little sense: some taxa may not have been seen, or there may not be a fixed number of taxa (e.g. in an open system; Peet 1974). As an alternative, *richness* (S) can be used for the concept of taxa number (McIntosh 1967).

It is not always possible to ensure that all sample sizes are equal and the number of different taxa increases with sample size and sampling effort (Magurran 1988). Then, *rarefaction* ($E(S)$) is the number of taxa expected if all samples were of a standard size (i.e. taxa per fixed number of individuals). Rarefaction assumes that imbalances between taxa are due to sampling and not to differences in actual abundances.

The following richness measures are available for count data:

- `margalef` Margalef richness index.
- `menhinick` Menhinick richness index.
- `none` Returns the number of observed taxa/types.

Value

- `index_richness()`, `simulate_richness()` and `index_composition()` return a `DiversityIndex` object.
- `bootstrap_*`() and `jackknife_*`() return a `data.frame`.

If `simplify` is `FALSE`, then `rarefaction()` returns a `list` (default), else return a `matrix`.

Asymptotic Species Richness

The following measures are available for count data:

`ace` Abundance-based Coverage Estimator.
`chao1` (improved/unbiased) Chao1 estimator.

The following measures are available for replicated incidence data:

`ice` Incidence-based Coverage Estimator.
`chao2` (improved/unbiased) Chao2 estimator.

Author(s)

N. Frerebeau

References

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See Also

[plot_diversity\(\)](#)

Other diversity: [heterogeneity-index](#), [similarity\(\)](#), [turnover-index](#)

Examples

```
## Richness
## Margalef and Menhinick index
## Data from Magurran 1988, p. 128-129
trap <- CountMatrix(data = c(9, 3, 0, 4, 2, 1, 1, 0, 1, 0, 1, 1,
                           1, 0, 1, 0, 0, 0, 1, 2, 0, 5, 3, 0),
                        nrow = 2, byrow = TRUE, dimnames = list(c("A", "B"), NULL))
index_richness(trap, method = "margalef") # 2.55 1.88
index_richness(trap, method = "menhinick") # 1.95 1.66

## Asymptotic species richness
## Chao1-type estimators
## Data from Chao & Chiu 2016
brazil <- CountMatrix(
  data = rep(x = c(1:21, 23, 25, 27, 28, 30, 32, 34:37, 41,
               45, 46, 49, 52, 89, 110, 123, 140),
             times = c(113, 50, 39, 29, 15, 11, 13, 5, 6, 6, 3, 4,
                       3, 5, 2, 5, 2, 2, 2, 1, 2, 1, 1, 1, 1, 1, 1,
                       0, 0, 2, 1, 1, 1, 1, 0, 1, 1, 1, 0, 0)),
  nrow = 1, byrow = TRUE
)

index_composition(brazil, method = c("chao1"), unbiased = FALSE) # 461.625
index_composition(brazil, method = c("ace"), k = 10) # 445.822

## Rarefaction
rarefaction(trap, sample = 13) # 6.56
```

Description

- `seriate_*`() computes a permutation order for rows and/or columns.
- `permute()` rearranges a data matrix according to a permutation order.
- `get_order()` returns the seriation order for rows and columns.

Usage

```

seriate_average(object, ...)

seriate_rank(object, ...)

permute(object, order, ...)

refine_seriation(object, ...)

## S4 method for signature 'PermutationOrder'
get_order(x)

## S4 method for signature 'CountMatrix,PermutationOrder'
permute(object, order)

## S4 method for signature 'IncidenceMatrix,PermutationOrder'
permute(object, order)

## S4 method for signature 'CA'
refine_seriation(
  object,
  cutoff,
  n = 1000,
  axes = c(1, 2),
  progress = getOption("tabula.progress"),
  ...
)

## S4 method for signature 'CountMatrix'
seriate_average(object, margin = c(1, 2), axes = 1, ...)

## S4 method for signature 'IncidenceMatrix'
seriate_average(object, margin = c(1, 2), axes = 1, ...)

## S4 method for signature 'CountMatrix'
seriate_rank(object, EPPM = FALSE, margin = c(1, 2), stop = 100)

## S4 method for signature 'IncidenceMatrix'
seriate_rank(object, margin = c(1, 2), stop = 100)

```

Arguments

object, x	An $m \times p$ data matrix (typically an object of class CountMatrix or IncidenceMatrix).
...	Further arguments to be passed to internal methods.
order	A PermutationOrder object giving the permutation order for rows and columns.
cutoff	A function that takes a numeric vector as argument and returns a single numeric value (see below).

n	A non-negative <code>integer</code> giving the number of bootstrap replications.
axes	An <code>integer</code> vector giving the subscripts of the CA axes to be used.
progress	A <code>logical</code> scalar: should a progress bar be displayed?
margin	A <code>numeric</code> vector giving the subscripts which the rearrangement will be applied over: 1 indicates rows, 2 indicates columns, <code>c(1, 2)</code> indicates rows then columns, <code>c(2, 1)</code> indicates columns then rows.
EPPM	A <code>logical</code> scalar: should the seriation be computed on EPPM instead of raw data?
stop	An <code>integer</code> giving the stopping rule (i.e. maximum number of iterations) to avoid infinite loop.

Value

- `seriate_*`() returns a `PermutationOrder` object.
- `refine_seriation()` returns a `RefineCA` object.
- `permute()` returns either a permuted `CountMatrix` or an `IncidenceMatrix` (the same as object).

Seriation

The matrix seriation problem in archaeology is based on three conditions and two assumptions, which Dunell (1970) summarizes as follows.

The homogeneity conditions state that all the groups included in a seriation must:

1. Be of comparable duration.
2. Belong to the same cultural tradition.
3. Come from the same local area.

The mathematical assumptions state that the distribution of any historical or temporal class:

1. Is continuous through time.
2. Exhibits the form of a unimodal curve.

These assumptions create a distributional model and ordering is accomplished by arranging the matrix so that the class distributions approximate the required pattern. The resulting order is inferred to be chronological.

The following seriation methods are available:

`seriate_average()` Correspondence analysis-based seriation (average ranking). Correspondence analysis (CA) is an effective method for the seriation of archaeological assemblages. The order of the rows and columns is given by the coordinates along one dimension of the CA space, assumed to account for temporal variation. The direction of temporal change within the correspondence analysis space is arbitrary: additional information is needed to determine the actual order in time.

`seriate_rank()` Reciprocal ranking seriation. These procedures iteratively rearrange rows and/or columns according to their weighted rank in the data matrix until convergence. Note that this procedure could enter into an infinite loop. If no convergence is reached before the maximum number of iterations, it stops with a warning.

Correspondence Analysis

`refine_seriation()` allows to identify samples that are subject to sampling error or samples that have underlying structural relationships and might be influencing the ordering along the CA space.

This relies on a partial bootstrap approach to CA-based seriation where each sample is replicated n times. The maximum dimension length of the convex hull around the sample point cloud allows to remove samples for a given cutoff value.

According to Peebles and Schachner (2012), "[this] point removal procedure [results in] a reduced dataset where the position of individuals within the CA are highly stable and which produces an ordering consistent with the assumptions of frequency seriation."

Note

Refining method can lead to much longer execution times and larger output objects.

Author(s)

N. Frerebeau

References

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See Also

[dimensio::ca\(\)](#)

Examples

```
## Replicates Desachy 2004 results
## Coerce dataset to abundance matrix
data("compiegne", package = "folio")
compiegne_count <- as_count(compiegne)

## Get seriation order for columns on EPPM using the reciprocal averaging method
## Expected column order: N, A, C, K, P, L, B, E, I, M, D, G, O, J, F, H
(compiegne_indices <- seriate_rank(compiegne_count, EPPM = TRUE, margin = 2))

## Permute columns
compiegne_new <- permute(compiegne_count, compiegne_indices)
```

```

## Plot new matrix
plot_ford(compiegne_new, EPPM = FALSE)

## See the vignette:
## Not run:
utils::vignette("seriation")

## End(Not run)

```

similarity	<i>Similarity</i>
------------	-------------------

Description

Similarity

Usage

```

similarity(object, ...)

## S4 method for signature 'CountMatrix'
similarity(
  object,
  method = c("brainerd", "bray", "jaccard", "morisita", "sorenson", "binomial"),
  ...
)

## S4 method for signature 'IncidenceMatrix'
similarity(object, method = c("jaccard", "sorenson"), ...)

```

Arguments

- object** A $m \times p$ matrix of count data.
- ...** Further arguments to be passed to internal methods.
- method** A **character** string specifying the method to be used (see details). Any unambiguous substring can be given.

Details

β -diversity can be measured by addressing *similarity* between pairs of samples/cases (Brainerd-Robinson, Jaccard, Morisita-Horn and Sorenson indices). Similarity between pairs of taxa/types can be measured by assessing the degree of co-occurrence (binomial co-occurrence).

Jaccard, Morisita-Horn and Sorenson indices provide a scale of similarity from 0-1 where 1 is perfect similarity and 0 is no similarity. The Brainerd-Robinson index is scaled between 0 and 200. The Binomial co-occurrence assessment approximates a Z-score.

binomial Binomial co-occurrence assessment. This assesses the degree of co-occurrence between taxa/types within a dataset. The strongest associations are shown by large positive numbers, the strongest segregations by large negative numbers.

brainerd Brainerd-Robinson quantitative index. This is a city-block metric of similarity between pairs of samples/cases.

bray Sorenson quantitative index (Bray and Curtis modified version of the Sorenson index).

jaccard Jaccard qualitative index.

morisita Morisita-Horn quantitative index.

sorenson Sorenson qualitative index.

Value

`similarity()` returns a [stats::dist](#) object.

Author(s)

N. Frerebeau

References

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See Also

Other diversity: [heterogeneity-index](#), [richness-index](#), [turnover-index](#)

Examples

```
## Data from Huntley 2008
ceramics <- CountMatrix(
  data = c(16, 9, 3, 0, 1,
         13, 3, 2, 0, 0,
         9, 5, 2, 5, 0,
         14, 12, 3, 0, 0,
         0, 26, 4, 0, 0,
         1, 26, 4, 0, 0,
         0, 11, 3, 13, 0,
         0, 0, 17, 0, 16,
```

```

    0, 0, 18, 0, 14),
nrow = 9, byrow = TRUE
)
rownames(ceramics) <- c("Atsinna", "Cienega", "Mirabal", "PdMuertos",
                         "Hesh", "LowPesc", "BoxS", "Ojo Bon", "S170")
colnames(ceramics) <- c("DLH-1", "DLH-2a", "DLH-2b", "DLH-2c", "DLH-4")

## Brainerd-Robinson measure (count data)
C <- similarity(ceramics, "brainerd")
plot_spot(C)

## Data from Magurran 1988, p. 166
data("birds", package = "folio")

## Plot spot diagram
birds <- as_count(birds)

## Jaccard measure (presence/absence data)
similarity(birds, "jaccard") # 0.46

## Sorenson measure (presence/absence data)
similarity(birds, "sorenson") # 0.63

## Jaccard measure (Bray's formula ; count data)
similarity(birds, "bray") # 0.44

## Morisita-Horn measure (count data)
similarity(birds, "morisita") # 0.81

```

subset*Extract or Replace Parts of an Object***Description**

Operators acting on objects to extract or replace parts.

Usage

```

## S4 method for signature 'DiversityIndex,ANY,missing'
x[[i]]

## S4 method for signature 'RefineCA,ANY,missing'
x[[i]]

## S4 method for signature 'PermutationOrder,ANY,missing'
x[[i]]

```

Arguments

- x An object from which to extract element(s) or in which to replace element(s).
- i A **character** string specifying elements to extract. Any unambiguous substring can be given (see details).

Value

A subsetted object.

Author(s)

N. Frerebeau

See Also

Other mutator: [mutator](#)

test_diversity *Diversity Test*

Description

Compares Shannon diversity between samples.

Usage

```
test_diversity(object, ...)

## S4 method for signature 'CountMatrix'
test_diversity(object, adjust = "holm", ...)
```

Arguments

- object A $m \times p$ matrix of count data.
- ... Further arguments to be passed to internal methods.
- adjust A **character** string specifying the method for adjusting p values (see [stats:::p.adjust\(\)](#)).

Details

This test produces two sided pairwise comparisons: it returns a matrix of adjusted p values.

Value

A **numeric matrix**.

Author(s)

N. Frerebeau

References

Magurran, A. E. (1988). *Ecological Diversity and its Measurement*. Princeton, NJ: Princeton University Press. doi: [10.1007/9789401573580](https://doi.org/10.1007/9789401573580).

See Also

Other statistics: [independance](#)

Examples

```
## Shannon diversity test
data("merzbach", package = "folio")
merzbach_count <- as_count(merzbach)
div <- test_diversity(merzbach_count)
```

turnover-index

Turnover

Description

Returns the degree of turnover in taxa composition along a gradient or transect.

Usage

```
turnover(object, ...)

## S4 method for signature 'CountMatrix'
turnover(
  object,
  method = c("whittaker", "cody", "routledge1", "routledge2", "routledge3", "wilson"),
  simplify = FALSE,
  ...
)

## S4 method for signature 'IncidenceMatrix'
turnover(
  object,
  method = c("whittaker", "cody", "routledge1", "routledge2", "routledge3", "wilson"),
  simplify = FALSE,
  ...
)
```

Arguments

object	A $m \times p$ matrix of count data.
...	Further arguments to be passed to internal methods.
method	A character string specifying the method to be used (see details). Any unambiguous substring can be given.
simplify	A logical scalar: should the result be simplified to a matrix?

Details

The following methods can be used to ascertain the degree of *turnover* in taxa composition along a gradient (β -diversity) on qualitative (presence/absence) data. This assumes that the order of the matrix rows (from 1 to n) follows the progression along the gradient/transect.

`whittaker` Whittaker measure.

`cody` Cody measure.

`routledge1` Routledge first measure.

`routledge2` Routledge second measure.

`routledge3` Routledge third measure. This is the exponential form of the second measure.

`wilson` Wilson measure.

Value

If `simplify` is FALSE, returns a `list` (default), else returns a `matrix`.

Author(s)

N. Frerebeau

References

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See Also

Other diversity: `heterogeneity-index`, `richness-index`, `similarity()`

Examples

```
## Data from Magurran 1988, p. 162
trees <- IncidenceMatrix(
  data = c(1, 1, 1, 0, 0, 0,
          1, 1, 1, 1, 1, 1,
          0, 0, 1, 0, 1, 0,
          0, 0, 0, 1, 1, 1,
          0, 0, 0, 0, 1, 1,
          0, 0, 0, 1, 0, 1),
  nrow = 6, byrow = FALSE,
  dimnames = list(c("1", "2", "3", "4", "5", "6")),
```

```
c("Birch", "Oak", "Rowan", "Beech", "Hazel", "Holly"))
)

## Whittaker's measure
turnover(trees, "whittaker") # 1

## Cody's measure
turnover(trees, "cody") # 3

## Routledge's measures
turnover(trees, method = c("routledge1", "routledge2", "routledge3"),
         simplify = TRUE) ## 0.29 0.56 1.75

## Wilson and Shmida's measure
turnover(trees, "wilson") # 1
```

Index

* **class**
 DiversityIndex, 2
 PermutationOrder-class, 10
 RefineCA-class, 17

* **diversity**
 heterogeneity-index, 3
 richness-index, 18
 similarity, 26
 turnover-index, 30

* **mutator**
 mutator, 9
 subset, 28

* **plot**
 plot_bar, 11
 plot_diversity, 13
 plot_line, 14
 plot_matrix, 15
 plot_spot, 16

* **seriation**
 seriation, 22

* **statistics**
 independance, 8
 test_diversity, 29

..., 11

.CompositionIndex (DiversityIndex), 2
.DiversityIndex (DiversityIndex), 2
.EvennessIndex (DiversityIndex), 2
.HeterogeneityIndex (DiversityIndex), 2
.PermutationOrder
 (PermutationOrder-class), 10
.RefineCA (RefineCA-class), 17
.RichnessIndex (DiversityIndex), 2
[[, DiversityIndex, ANY, missing-method
 (subset), 28
[[, PermutationOrder, ANY, missing-method
 (subset), 28
[[, RefineCA, ANY, missing-method
 (subset), 28

Bertin (plot_bar), 11

Bertin diagram, 17

bootstrap_evenness
 (heterogeneity-index), 3

bootstrap_evenness, CountMatrix-method
 (heterogeneity-index), 3

bootstrap_evenness-method
 (heterogeneity-index), 3

bootstrap_heterogeneity
 (heterogeneity-index), 3

bootstrap_heterogeneity, CountMatrix-method
 (heterogeneity-index), 3

bootstrap_heterogeneity-method
 (heterogeneity-index), 3

bootstrap_richness (richness-index), 18

bootstrap_richness, CountMatrix-method
 (richness-index), 18

bootstrap_richness-method
 (richness-index), 18

character, 3, 5, 10, 14, 18, 20, 26, 29, 30

CompositionIndex-class
 (DiversityIndex), 2

CountMatrix, 5, 8, 23, 24

data.frame, 6, 21

dimensio::CA, 18

dimensio::ca(), 25

DiversityIndex, 2, 6, 10, 13, 18, 21

DiversityIndex-class (DiversityIndex), 2

eppm (independance), 8

eppm(), 12

eppm, CountMatrix-method (independance),
 8

eppm-method (independance), 8

EvennessIndex-class (DiversityIndex), 2

Ford (plot_bar), 11

function, 11, 16

get (mutator), 9

get_index (mutator), 9
 get_index, EvennessIndex-method
 (mutator), 9
 get_index, HeterogeneityIndex-method
 (mutator), 9
 get_index, RichnessIndex-method
 (mutator), 9
 get_index-method (mutator), 9
 get_method (mutator), 9
 get_method, DiversityIndex-method
 (mutator), 9
 get_method-method (mutator), 9
 get_order (mutator), 9
 get_order, PermutationOrder-method
 (seriation), 22
 get_order-method (mutator), 9
 ggplot2::ggplot, 11, 14, 15, 17

heterogeneity-index, 3
 HeterogeneityIndex-class
 (DiversityIndex), 2

IncidenceMatrix, 23, 24
 independance, 8, 30
 index_composition (richness-index), 18
 index_composition, CountMatrix-method
 (richness-index), 18
 index_composition, IncidenceMatrix-method
 (richness-index), 18
 index_composition-method
 (richness-index), 18
 index_evenness (heterogeneity-index), 3
 index_evenness(), 13
 index_evenness, CountMatrix-method
 (heterogeneity-index), 3
 index_evenness-method
 (heterogeneity-index), 3
 index_heterogeneity
 (heterogeneity-index), 3
 index_heterogeneity(), 13
 index_heterogeneity, CountMatrix-method
 (heterogeneity-index), 3
 index_heterogeneity-method
 (heterogeneity-index), 3
 index_richness (richness-index), 18
 index_richness(), 13
 index_richness, CountMatrix-method
 (richness-index), 18

index_richness-method (richness-index),
 18
 integer, 3, 5, 10, 17, 18, 20, 24

jackknife_evenness
 (heterogeneity-index), 3
 jackknife_evenness, CountMatrix-method
 (heterogeneity-index), 3
 jackknife_evenness-method
 (heterogeneity-index), 3
 jackknife_heterogeneity
 (heterogeneity-index), 3
 jackknife_heterogeneity, CountMatrix-method
 (heterogeneity-index), 3
 jackknife_heterogeneity-method
 (heterogeneity-index), 3
 jackknife_richness (richness-index), 18
 jackknife_richness, CountMatrix-method
 (richness-index), 18
 jackknife_richness-method
 (richness-index), 18

list, 21, 31
 logical, 5, 11, 14, 16, 20, 24, 30

matrigraphe (plot_matrix), 15
 matrix, 8, 21, 29, 31
 mutator, 9, 29

numeric, 3, 5, 8, 17, 18, 20, 24, 29

PermutationOrder, 23, 24
 PermutationOrder-class, 10
 permute (seriation), 22
 permute, CountMatrix, PermutationOrder-method
 (seriation), 22
 permute, IncidenceMatrix, PermutationOrder-method
 (seriation), 22
 permute-method (seriation), 22
 plot, DiversityIndex, missing-method
 (plot_diversity), 13
 plot_bar, 11, 13–15, 17
 plot_bertin (plot_bar), 11
 plot_bertin, matrix-method (plot_bar), 11
 plot_bertin-method (plot_bar), 11
 plot_diversity, 12, 13, 14, 15, 17
 plot_diversity(), 7, 22
 plot_ford (plot_bar), 11
 plot_ford(), 9

```

plot_ford,CountMatrix-method
    (plot_bar), 11
plot_ford,matrix-method (plot_bar), 11
plot_ford-method (plot_bar), 11
plot_heatmap (plot_matrix), 15
plot_heatmap(), 9
plot_heatmap,matrix-method
    (plot_matrix), 15
plot_heatmap-method (plot_matrix), 15
plot_line, 12, 13, 14, 15, 17
plot_matrix, 12–14, 15, 17
plot_rank (plot_line), 14
plot_rank,matrix-method (plot_line), 14
plot_rank-method (plot_line), 14
plot_spot, 12–15, 16
plot_spot,dist-method (plot_spot), 16
plot_spot,matrix-method (plot_spot), 16
plot_spot,OccurrenceMatrix-method
    (plot_spot), 16
plot_spot-method (plot_spot), 16
pvi (independance), 8
pvi(), 15
pvi,CountMatrix-method (independance), 8
pvi-method (independance), 8

rarefaction (richness-index), 18
rarefaction,CountMatrix-method
    (richness-index), 18
rarefaction-method (richness-index), 18
refine_seriation (seriation), 22
refine_seriation,CA-method (seriation),
    22
refine_seriation-method (seriation), 22
RefineCA, 24
RefineCA-class, 17
richness-index, 18
RichnessIndex-class (DiversityIndex), 2

seriate_average (seriation), 22
seriate_average,CountMatrix-method
    (seriation), 22
seriate_average,IncidenceMatrix-method
    (seriation), 22
seriate_average-method (seriation), 22
seriate_rank (seriation), 22
seriate_rank(), 9
seriate_rank,CountMatrix-method
    (seriation), 22
seriate_rank,IncidenceMatrix-method
    (seriation), 22
seriate_rank-method (seriation), 22
seriation, 22
seriographe (plot_bar), 11
set (mutator), 9
similarity, 7, 22, 26, 31
similarity(), 7
similarity,CountMatrix-method
    (similarity), 26
similarity,IncidenceMatrix-method
    (similarity), 26
similarity-method (similarity), 26
simulate_evenness
    (heterogeneity-index), 3
simulate_evenness,CountMatrix-method
    (heterogeneity-index), 3
simulate_evenness-method
    (heterogeneity-index), 3
simulate_heterogeneity
    (heterogeneity-index), 3
simulate_heterogeneity,CountMatrix-method
    (heterogeneity-index), 3
simulate_heterogeneity-method
    (heterogeneity-index), 3
simulate_richness (richness-index), 18
simulate_richness,CountMatrix-method
    (richness-index), 18
simulate_richness-method
    (richness-index), 18
stats::dist, 27
stats::p.adjust(), 29
stats::quantile(), 5, 20
subset, 10, 28

test_diversity, 9, 29
test_diversity,CountMatrix-method
    (test_diversity), 29
test_diversity-method (test_diversity),
    29
turnover (turnover-index), 30
turnover(), 7
turnover,CountMatrix-method
    (turnover-index), 30
turnover,IncidenceMatrix-method
    (turnover-index), 30
turnover-index, 30
turnover-method (turnover-index), 30

```